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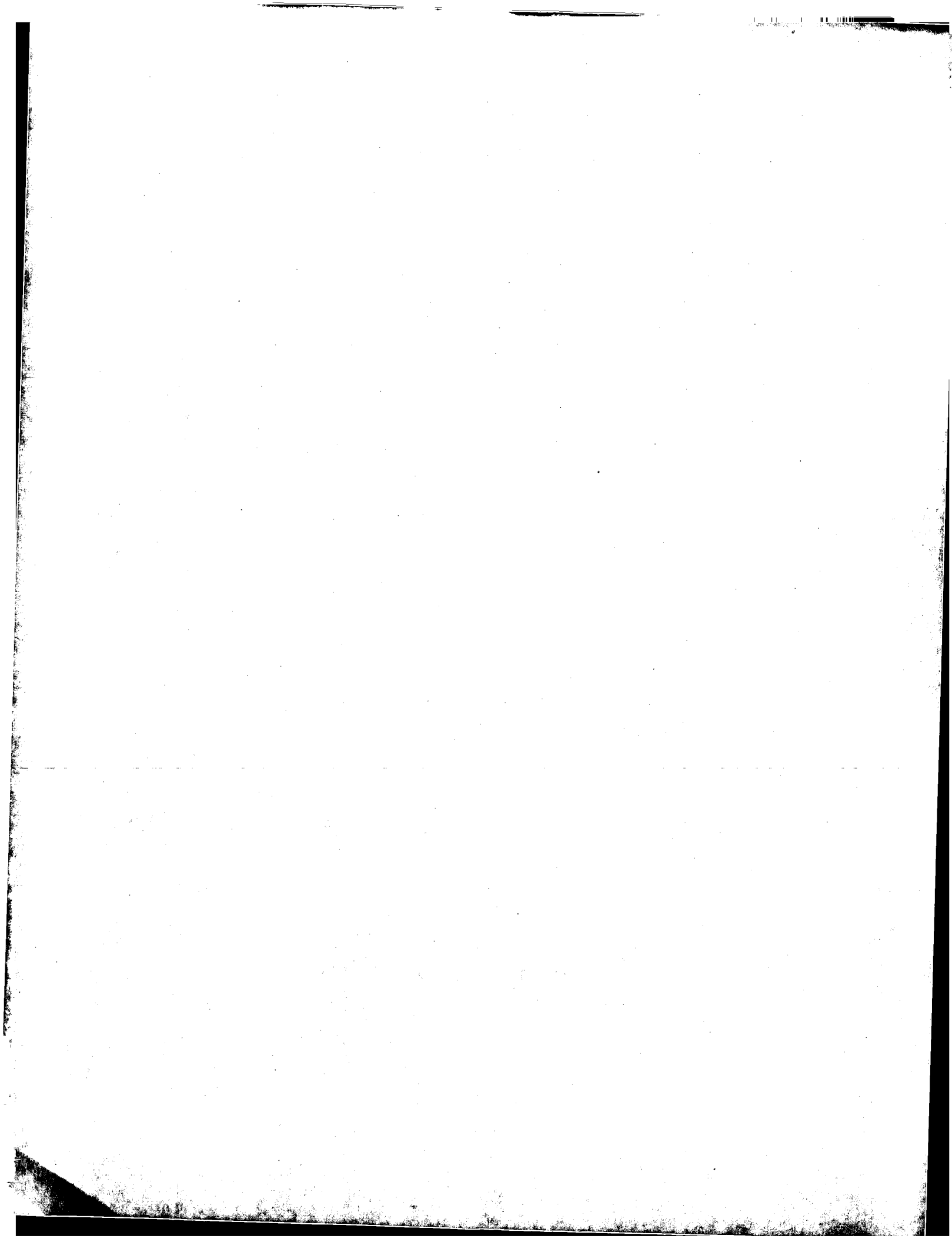
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(54) Title: DSP-15 DUAL-SPECIFICITY PHOSPHATASE

MALVTVSRSPPGSGASTPVGPWDQAVQRRSRLQRRQSFVLRGAVLGLQDGGDNDAAEASSEPTEKAPSEELHGD  
 QTDGQGSQSPQKEEQROHLHLMVQLLRPQDDIRLAAQLEAPRPPRLRYLLVSTREGEGLSQDETLLGVDFPDS  
 SSPSCITGLVLPWSDTQVYLDGDDGFSVTSGGQSRIFKPIISIQTMWATLQVLHQACEAALGSGLPVGGGSLTWASH  
 YQERLNSEQSCLNEWAMADLESLRPPSAEPGSGSEQMEQATRAELWKVLDVSDLESVTSKEIRQALELRGLPL  
 QQYRDFIDNQMLLLVAQRDRASRIFPHLYLGSEWNAANLEELQRNRVTHILNMAREIDNFYPERFTHYHVRLLWDEES  
 AQLLPHWKETHRFIEAARAQGTHTLVHCKMGVSRSAATVLAAMKQYECSEALRHVQELRPIARPNGFLRLQIQI  
 YQGILTASRQSHVWEQVGGVSPPEHPAPEVSTPFPPLPPEPEGGGEEKVVGMEESQAAPKEEPPRPRINLRGVMR  
 SISLLEPSLELESTSETSDMPEVFSHESHSHEEPLQFPPLARTKGGQVDRGPQALKSRQSVVTLQGSVVANRT  
 QAFQEQQGQGGQGEPCISSTPRFRKVVQASVHDSGEEGEA

(57) Abstract: Compositions and methods are provided for the treatment of conditions associated with cell proliferation, cell differentiation and cell survival. In particular, the dual-specificity phosphatase DSP-15, and polypeptide variants thereof that stimulate dephosphorylation of DSP-15 substrates, are provided. The polypeptides may be used, for example, to identify antibodies and other agents that inhibit DSP-15 activity. The polypeptides and agents may be used to modulate cell proliferation, differentiation and survival.

WO 02/24740 A2

## DSP-15 DUAL-SPECIFICITY PHOSPHATASE

## TECHNICAL FIELD

The present invention relates generally to compositions and methods useful for treating conditions associated with defects in cell proliferation, cell differentiation and/or cell survival. The invention is more particularly related to dual-specificity protein phosphatases, and polypeptide variants thereof. The present invention is also related to the use of such polypeptides to identify antibodies and other agents, including small molecules, that modulate signal transduction leading to proliferative responses, cell differentiation and/or cell survival.

## 10 BACKGROUND OF THE INVENTION

Mitogen-activated protein kinases (MAP-kinases) are present as components of conserved cellular signal transduction pathways that have a variety of conserved members. MAP-kinases are activated by phosphorylation at a dual phosphorylation motif with the sequence Thr-X-Tyr (by MAP-kinase kinases), in which phosphorylation at the tyrosine and threonine residues is required for activity. Activated MAP-kinases phosphorylate several transduction targets, including transcription factors. Inactivation of MAP-kinases is mediated by dephosphorylation at this site by dual-specificity phosphatases referred to as MAP-kinase phosphatases. In higher eukaryotes, the physiological role of MAP-kinase signaling has been correlated with cellular events such as proliferation, oncogenesis, development and differentiation. Accordingly, the ability to regulate signal transduction via these pathways could lead to the development of treatments and preventive therapies for human diseases associated with MAP-kinase signaling, such as cancer.

Dual-specificity protein tyrosine phosphatases (dual-specificity phosphatases) are phosphatases that dephosphorylate both phosphotyrosine and phosphothreonine/serine residues (Walton et al., *Ann. Rev. Biochem.* 62:101-120, 1993). Several dual-specificity phosphatases that inactivate a MAP-kinase have been identified, including MKP-1 (WO 97/00315; Keyse and Emslie, *Nature* 59:644-647, 1992), MKP-2 (WO97/00315), MKP-4, MKP-5, MKP-7, Hb5 (WO 97/06245), PAC1

(Ward et al., *Nature* 367:651-654, 1994), HVH2 (Guan and Butch, *J. Biol. Chem.* 270:7197-7203, 1995) and PYST1 (Groom et al., *EMBO J.* 15:3621-3632, 1996). Expression of certain dual-specificity phosphatases is induced by stress or mitogens, but others appear to be expressed constitutively in specific cell types. The regulation of  
5 dual-specificity phosphatase expression and activity is critical for control of MAP-kinase mediated cellular functions, including cell proliferation, cell differentiation and cell survival. For example, dual-specificity phosphatases may function as negative regulators of cell proliferation. It is likely that there are many such dual-specificity phosphatases, with varying specificity with regard to cell type or activation. However,  
10 the regulation of dual specificity phosphatases remains poorly understood and only a relatively small number of dual-specificity phosphatases have been identified.

Accordingly, there is a need in the art for an improved understanding of MAP-kinase signaling, and the regulation of dual-specificity phosphatases within MAP-kinase signaling cascades. An increased understanding of dual-specificity phosphatase  
15 regulation may facilitate the development of methods for modulating the activity of proteins involved in MAP-kinase cascades, and for treating conditions associated with such cascades. The present invention fulfills these needs and further provides other related advantages.

#### SUMMARY OF THE INVENTION

20 Briefly stated, the present invention provides compositions and methods for identifying agents capable of modulating cellular proliferative responses. In one aspect, the present invention provides isolated DSP-15 polypeptides having the sequence of DSP-15 recited in SEQ ID NO:2, or a variant thereof that differs in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of  
25 the residues in SEQ ID NO:2, such that the polypeptide retains the ability to dephosphorylate an activated MAP-kinase.

Within further aspects, the present invention provides an isolated polynucleotide that encodes at least ten consecutive amino acids of a polypeptide having a sequence corresponding to SEQ ID NO:2. In certain embodiments the invention  
30 provides an isolated polynucleotide that encodes at least fifteen consecutive amino acids

of a polypeptide having a sequence corresponding to SEQ ID NO:2. Certain such polynucleotides encode a DSP-15 polypeptide. Still further, polynucleotides may be antisense polynucleotides that comprise at least 15 consecutive nucleotides complementary to a portion of a DSP-15 polynucleotide and/or that detectably hybridize to the complement of the sequence recited in SEQ ID NO:1 under conditions that include a wash in 0.1X SSC and 0.1% SDS at 50°C for 15 minutes. Also provided are expression vectors comprising any of the foregoing polynucleotides, and host cells transformed or transfected with such expression vectors.

The present invention further provides, within other aspects, methods for producing a DSP-15 polypeptide, comprising the steps of: (a) culturing a host cell as described above under conditions that permit expression of the DSP-15 polypeptide; and (b) isolating DSP-15 polypeptide from the host cell culture.

Also provided by the present invention are isolated antibodies, and antigen binding fragments thereof, that specifically bind to a DSP-15 polypeptide such as a polypeptide having the sequence of SEQ ID NO:2.

The present invention further provides, within other aspects, pharmaceutical compositions comprising a polypeptide, polynucleotide, antibody or fragment thereof as described above in combination with a physiologically acceptable carrier.

Within further aspects, the present invention provides methods for detecting DSP-15 expression in a sample, comprising: (a) contacting a sample with an antibody or an antigen-binding fragment thereof as described above, under conditions and for a time sufficient to allow formation of an antibody/DSP-15 complex; and (b) detecting the level of antibody/DSP-15 complex.

Within still other aspects, the present invention provides methods for detecting DSP-15 expression in a sample, comprising: (a) contacting a sample with an antisense polynucleotide as described above; and (b) detecting in the sample an amount of DSP-15 polynucleotide that hybridizes to the antisense polynucleotide. The amount of DSP-15 polynucleotide that hybridizes to the antisense polynucleotide may be determined, for example, using polymerase chain reaction or a hybridization assay.

The invention also provides DSP-15 polypeptides useful in screening assays for modulators of enzyme activity and/or substrate binding. Methods are also provided, within other aspects, for screening for an agent that modulates DSP-15 activity, comprising the steps of: (a) contacting a candidate agent with a DSP-15 polypeptide as described above, under conditions and for a time sufficient to permit interaction between the polypeptide and candidate agent; and (b) subsequently evaluating the ability of the polypeptide to dephosphorylate a DSP-15 substrate, relative to a predetermined ability of the polypeptide to dephosphorylate the DSP-15 substrate in the absence of candidate agent. Such methods may be performed *in vitro* or in a cellular environment (*e.g.*, within an intact cell).

Within further aspects, methods are provided for screening for an agent that modulates DSP-15 activity, comprising the steps of: (a) contacting a candidate agent with a cell comprising a DSP-15 promoter operably linked to a polynucleotide encoding a detectable transcript or protein, under conditions and for a time sufficient to permit interaction between the promoter and candidate agent; and (b) subsequently evaluating the expression of the polynucleotide, relative to a predetermined level of expression in the absence of candidate agent.

Also provided are methods for modulating a proliferative response in a cell, comprising contacting a cell with an agent that modulates DSP-15 activity.

Within further aspects, methods are provided for modulating differentiation of a cell, comprising contacting a cell with an agent that modulates DSP-15 activity.

The present invention further provides methods for modulating cell survival, comprising contacting a cell with an agent that modulates DSP-15 activity.

Within related aspects, the present invention provides methods for treating a patient afflicted with a disorder associated with DSP-15 activity (or treatable by administration of DSP-15), comprising administering to a patient a therapeutically effective amount of an agent that modulates DSP-15 activity. Such disorders include Duchenne Muscular Dystrophy, as well as cancer, graft-versus-host disease, autoimmune diseases, allergies, metabolic diseases, abnormal cell growth, abnormal cell proliferation and cell cycle abnormalities.

Within further aspects, DSP-15 substrate trapping mutant polypeptides are provided. Such polypeptides differ from the sequence recited in SEQ ID NO:2 in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of the residues in SEQ ID NO:2, such that the polypeptide binds to a substrate with an affinity that is not substantially diminished relative to DSP-15, and such that the ability of the polypeptide to dephosphorylate a substrate is reduced relative to DSP-15. Within certain specific embodiments, a substrate trapping mutant polypeptide contains a substitution at position 382 or position 413 of SEQ ID NO:2.

The present invention further provides, within other aspects, methods for screening a molecule for the ability to interact with DSP-15, comprising the steps of: (a) contacting a candidate molecule with a polypeptide as described above under conditions and for a time sufficient to permit the candidate molecule and polypeptide to interact; and (b) detecting the presence or absence of binding of the candidate molecule to the polypeptide. The step of detecting may comprise, for example, an affinity purification step, a yeast two hybrid screen or a screen of a phage display library.

In one aspect, the present invention provides isolated DSP-15 polypeptides comprising the sequence of DSP-15 alternate form recited in SEQ ID NO:21, or a variant thereof that differs in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of the residues in SEQ ID NO:21, such that the polypeptide retains the ability to dephosphorylate an activated MAP-kinase.

Within further aspects, the present invention provides an isolated polynucleotide that encodes at least ten consecutive amino acids of a polypeptide having a sequence corresponding to SEQ ID NO:21. In certain embodiments the invention provides an isolated polynucleotide that encodes at least fifteen consecutive amino acids of a polypeptide having a sequence corresponding to SEQ ID NO:21. Certain such polynucleotides encode a DSP-15 alternate form polypeptide. Still further, polynucleotides may be antisense polynucleotides that comprise at least 15 consecutive nucleotides complementary to a portion of a DSP-15 alternate form polynucleotide and/or that detectably hybridize to the complement of the sequence recited in SEQ ID NO:20 under conditions that include a wash in 0.1X SSC and 0.1% SDS at 60°C for 15



minutes. Also provided are expression vectors comprising any of the foregoing polynucleotides, and host cells transformed or transfected with such expression vectors.

The present invention further provides, within other aspects, methods for producing a DSP-15 alternate form polypeptide, comprising the steps of: (a) culturing a  
5 host cell as described above under conditions that permit expression of the DSP-15 alternate form polypeptide; and (b) isolating DSP-15 alternate form polypeptide from the host cell culture.

Also provided by the present invention are isolated antibodies, and antigen binding fragments thereof, that specifically bind to a DSP-15 alternate form  
10 polypeptide such as a polypeptide having the sequence of SEQ ID NO:21.

The present invention further provides, within other aspects, pharmaceutical compositions comprising a polypeptide, polynucleotide, antibody or fragment thereof as described above in combination with a physiologically acceptable carrier.

15 Within further aspects, the present invention provides methods for detecting DSP-15 alternate form expression in a sample, comprising: (a) contacting a sample with an antibody or an antigen-binding fragment thereof as described above, under conditions and for a time sufficient to allow formation of an antibody/DSP-15 alternate form complex; and (b) detecting the level of antibody/DSP-15 alternate form  
20 complex.

Within still other aspects, the present invention provides methods for detecting DSP-15 alternate form expression in a sample, comprising: (a) contacting a sample with an antisense polynucleotide as described above; and (b) detecting in the sample an amount of DSP-15 alternate form polynucleotide that hybridizes to the  
25 antisense polynucleotide. The amount of DSP-15 alternate form polynucleotide that hybridizes to the antisense polynucleotide may be determined, for example, using polymerase chain reaction or a hybridization assay.

The invention also provides DSP-15 alternate form polypeptides useful in screening assays for modulators of enzyme activity and/or substrate binding.  
30 Methods are also provided, within other aspects, for screening for an agent that modulates DSP-15 alternate form activity, comprising the steps of: (a) contacting a

candidate agent with a polypeptide as described above, under conditions and for a time sufficient to permit interaction between the polypeptide and candidate agent; and (b) subsequently evaluating the ability of the polypeptide to dephosphorylate a DSP-15 alternate form substrate, relative to a predetermined ability of the polypeptide to dephosphorylate the DSP-15 alternate form substrate in the absence of candidate agent. Such methods may be performed *in vitro* or in a cellular environment (e.g., within an intact cell).

Within further aspects, methods are provided for screening for an agent that modulates DSP-15 alternate form activity, comprising the steps of: (a) contacting a candidate agent with a cell comprising a DSP-15 alternate form promoter operably linked to a polynucleotide encoding a detectable transcript or protein, under conditions and for a time sufficient to permit interaction between the promoter and candidate agent; and (b) subsequently evaluating the expression of the polynucleotide, relative to a predetermined level of expression in the absence of candidate agent.

Also provided are methods for modulating a proliferative response in a cell, comprising contacting a cell with an agent that modulates DSP-15 alternate form activity.

Within further aspects, methods are provided for modulating differentiation of a cell, comprising contacting a cell with an agent that modulates DSP-15 alternate form activity.

The present invention further provides methods for modulating cell survival, comprising contacting a cell with an agent that modulates DSP-15 alternate form activity.

Within related aspects, the present invention provides methods for treating a patient afflicted with a disorder associated with DSP-15 alternate form activity (or treatable by administration of DSP-15 alternate form), comprising administering to a patient a therapeutically effective amount of an agent that modulates DSP-15 alternate form activity. Such disorders include cancer, graft-versus-host disease, autoimmune diseases, allergies, metabolic diseases, abnormal cell growth, abnormal cell proliferation and cell cycle abnormalities.

Within further aspects, DSP-15 alternate form substrate trapping mutant polypeptides are provided. Such polypeptides differ from the sequence recited in SEQ ID NO:21 in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of the residues in SEQ ID NO:21, such that the polypeptide binds to  
5 a substrate with an affinity that is not substantially diminished relative to DSP-15 alternate form, and such that the ability of the polypeptide to dephosphorylate a substrate is reduced relative to DSP-15 alternate form. Within certain specific embodiments, a substrate trapping mutant polypeptide contains a substitution at position 382 or position 413 of SEQ ID NO:21.

10 The present invention further provides, within other aspects, methods for screening a molecule for the ability to interact with DSP-15 alternate form, comprising the steps of: (a) contacting a candidate molecule with a DSP-15 alternate form polypeptide or variant thereof as described above under conditions and for a time sufficient to permit the candidate molecule and polypeptide to interact; and (b) detecting  
15 the presence or absence of binding of the candidate molecule to the polypeptide. The step of detecting may comprise, for example, an affinity purification step, a yeast two hybrid screen or a screen of a phage display library.

These and other aspects of the present invention will become apparent  
20 upon reference to the following detailed description and attached drawings. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 presents a cDNA sequence for DSP-15 (SEQ ID NO:1), with  
25 the start and stop codons shown in bold.

Figure 2 presents the predicted amino acid sequence of DSP-15 (SEQ ID NO:2).

Figure 3 is a sequence alignment of GenBank Accession Number AK001790(SEQ ID NO:12) and several MAP-kinase phosphatases (SEQ ID Nos: 3-  
30 11).

Figure 4 shows a cDNA sequence for a murine DSP-15 variant (SEQ ID NO:20), with the start and stop codons shown in bold.

Figure 5 presents the predicted amino acid sequence of the murine DSP-15 variant (SEQ ID NO:21) encoded by the protein coding region of SEQ ID NO:20.

5           Figure 6 shows a sequence alignment of DSP-15 (SEQ ID NO:2) with other MAP kinase phosphatases (SEQ ID Nos: 13-15).

## DETAILED DESCRIPTION OF THE INVENTION

As noted above, the present invention is generally directed to compositions and methods for modulating (*i.e.*, stimulating or inhibiting) cellular proliferative responses, *in vitro* and *in vivo*. In particular, the present invention provides a dual-specificity phosphatase DSP-15 or DSP-15 alternate form (Figures 1-2, 4-5; SEQ ID NOs:1, 2, 20, 21), as well as variants thereof and antibodies that specifically bind DSP-15 or DSP-15 alternate form. Also provided herein are methods for using such compounds for screens, detection assays and related therapeutic uses.

### 15    DSP-15 POLYPEPTIDES AND POLYNUCLEOTIDES

As used herein, the term "DSP-15 polypeptide" or "DSP-15 alternate form polypeptide" refers to a polypeptide that comprises a DSP-15 sequence as provided herein or a variant of such a sequence. Such polypeptides are capable of dephosphorylating both tyrosine and threonine/serine residues in a DSP-15 substrate, with an activity that is not substantially diminished relative to that of a full length native DSP-15. DSP-15 substrates include activated (*i.e.*, phosphorylated) MAP-kinases. Other substrates may be identified using substrate trapping mutants, as described herein, and include polypeptides having one or more phosphorylated tyrosine, threonine and/or serine residues.

25           DSP-15 or DSP-15 alternate form polypeptide variants within the scope of the present invention may contain one or more substitutions, deletions, additions and/or insertions. For certain DSP-15 or DSP-15 alternate form variants, the ability of the variant to dephosphorylate tyrosine and threonine residues within a DSP-15 substrate is not substantially diminished. The ability of such a DSP-15 variant to

dephosphorylate tyrosine and threonine residues within a DSP-15 substrate may be enhanced or unchanged, relative to a native DSP-15 or DSP-15 alternate form, or may be diminished by less than 50%, and preferably less than 20%, relative to native DSP-15 or DSP-15 alternate form. Such variants may be identified using the representative  
5 assays provided herein.

Also contemplated by the present invention are modified forms of DSP-15 and/or DSP-15 alternate form in which a specific function is disabled. For example, such proteins may be constitutively active or inactive, or may display altered binding or catalytic properties. Such altered proteins may be generated using well known  
10 techniques, and the altered function confirmed using screens such as those provided herein. Certain modified DSP-15 or DSP-15 alternate form polypeptides are known as "substrate trapping mutants." Such polypeptides retain the ability to bind a substrate (*i.e.*,  $K_m$  is not substantially diminished), but display a reduced ability to dephosphorylate a substrate (*i.e.*,  $k_{cat}$  is reduced, preferably to less than 1 per minute).  
15 Further, the stability of the substrate trapping mutant/substrate complex should not be substantially diminished, relative to the stability of a DSP-15/substrate complex, including a DSP-5 alternate form/substrate complex. Complex stability may be assessed based on the association constant ( $K_a$ ). Determination of  $K_m$ ,  $k_{cat}$  and  $K_a$  may be readily accomplished using standard techniques known in the art (*see, e.g.*, WO  
20 98/04712; Lehninger, Biochemistry, 1975 Worth Publishers, NY) and assays provided herein. Substrate trapping mutants may be generated, for example, by modifying DSP-15 with an amino acid substitution at position 382 or position 413 (*e.g.*, by replacing the amino acid aspartate at position 382 with an alanine residue, or by replacing the cysteine at residue 413 with a serine). Substrate trapping mutants may be used, for example, to  
25 identify DSP-15 substrates. Briefly, the modified DSP-15 or DSP-15 alternate form may be contacted with a candidate substrate (alone or within a mixture of proteins, such as a cell extract) to permit the formation of a substrate/DSP-15 complex. The complex may then be isolated by conventional techniques to permit the isolation and characterization of substrate. The preparation and use of substrate trapping mutants is  
30 described, for example, within PCT Publication No. WO 98/04712.

Preferably, a variant contains conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydrophobic nature of the polypeptide to be substantially unchanged. Amino acid substitutions may generally be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine. Other groups of amino acids that may represent conservative changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his. A variant may also, or alternatively, contain nonconservative changes.

In general, modifications may be more readily made in non-critical regions, which are regions of the native sequence that do not substantially change the activity of DSP-15 or DSP-15 alternate form. Non-critical regions may be identified by modifying the DSP-15 sequence in a particular region and assaying the ability of the resulting variant in a phosphatase assay, as described herein. Preferred sequence modifications are made so as to retain the active site domain (VHCKMGVSRS, SEQ ID NO:16). Within certain preferred embodiments, such modifications affect interactions between DSP-15 (or DSP-15 alternate form) and cellular components other than DSP-15 substrates. However, substitutions may also be made in critical regions of the native protein, provided that the resulting variant substantially retains the ability to stimulate substrate dephosphorylation. Within certain embodiments, a variant contains substitutions, deletions, additions and/or insertions at no more than 50%, preferably no more than 25%, of the amino acid residues.

Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the activity of the polypeptide. In particular, variants may contain additional amino acid sequences at the

amino and/or carboxy termini. Such sequences may be used, for example, to facilitate purification or detection of the polypeptide.

DSP-15 (or DSP-15 alternate form) polypeptides may be prepared using any of a variety of well known techniques. Recombinant polypeptides encoded by DNA sequences as described below may be readily prepared from the DNA sequences using any of a variety of expression vectors known to those having ordinary skill in the art. Expression may be achieved in any appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast and higher eukaryotic cells (including mammalian cells), and forms that differ in glycosylation may be generated by varying the host cell or post-isolation processing. Supernatants from suitable host/vector systems which secrete recombinant protein or polypeptide into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion exchange resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant polypeptide.

Portions and other variants having fewer than about 100 amino acids, and generally fewer than about 50 amino acids, may also be generated by synthetic procedures, using techniques well known to those having ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. See Merrifield, *J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin-Elmer, Inc., Applied BioSystems Division (Foster City, CA), and may be operated according to the manufacturer's instructions.

A "DSP-15 polynucleotide" is any polynucleotide that encodes at least a portion of a DSP-15 or DSP-15 alternate form polypeptide or a variant thereof, or that is complementary to such a polynucleotide. Preferred polynucleotides comprise at least 15 consecutive nucleotides, preferably at least 30 consecutive nucleotides, that encode a DSP-15 or DSP-15 alternate form polypeptide or that are complementary to such a

sequence. Certain polynucleotides encode a DSP-15 or DSP-15 alternate form polypeptide; others may find use as probes, primers or antisense oligonucleotides, as described below. Polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA molecules.

5 Additional coding or non-coding sequences may, but need not, be present within a polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

DSP-15 polynucleotides may comprise a native sequence (*i.e.*, an endogenous DSP-15 or DSP-15 alternate form sequence, or a portion or splice variant thereof) or may comprise a variant of such a sequence. Polynucleotide variants may

10 contain one or more substitutions, additions, deletions and/or insertions such that the activity of the encoded polypeptide is not substantially diminished, as described above. The effect on the activity of the encoded polypeptide may generally be assessed as described herein. Variants preferably exhibit at least about 70% identity, more

15 preferably at least about 80% identity and most preferably at least about 90% identity to a polynucleotide sequence that encodes a native DSP-15 or DSP-15 alternate form or a portion thereof. The percent identity may be readily determined by comparing sequences using computer algorithms well known to those having ordinary skill in the art, such as Align or the BLAST algorithm (Altschul, *J. Mol. Biol.* 219:555-565, 1991;

20 Henikoff and Henikoff, *Proc. Natl. Acad. Sci. USA* 89:10915-10919, 1992), which is available at the NCBI website (<http://www.ncbi.nlm.nih.gov/cgi-bin/BLAST>). Default parameters may be used. Certain variants are substantially homologous to a native gene. Such polynucleotide variants are capable of hybridizing under moderately stringent conditions to a naturally occurring DNA or RNA sequence encoding a native

25 DSP-15 or DSP-15 alternate form (or a complementary sequence). Suitable moderately stringent conditions include, for example, prewashing in a solution of 5 X SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50°C-70°C, 5 X SSC, for 1-16 hours (*e.g.*, overnight); followed by washing once or twice at 22-65°C for 20-40 minutes with one or more each of 2X, 0.5X and 0.2X SSC containing 0.05-0.1% SDS. For additional

30 stringency, conditions may include a wash in 0.1X SSC and 0.1% SDS at 50-60 °C for 15-40 minutes. As known to those having ordinary skill in the art, variations in



stringency of hybridization conditions may be achieved by altering the time, temperature and/or concentration of the solutions used for prehybridization, hybridization and wash steps, and suitable conditions may also depend in part on the particular nucleotide sequences of the probe used, and of the blotted, proband nucleic acid sample. Accordingly, it will be appreciated that suitably stringent conditions can be readily selected without undue experimentation where a desired selectivity of the probe is identified, based on its ability to hybridize to one or more certain proband sequences while not hybridizing to certain other proband sequences.

It will also be appreciated by those having ordinary skill in the art that, as a result of the degeneracy of the genetic code, there are many nucleotide sequences that encode a polypeptide as described herein. Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention.

Polynucleotides may be prepared using any of a variety of techniques. For example, a polynucleotide may be amplified from cDNA prepared from a suitable cell or tissue type, such as human skeletal muscle cells. Such polynucleotides may be amplified via polymerase chain reaction (PCR). For this approach, sequence-specific primers may be designed based on the sequences provided herein, and may be purchased or synthesized.

An amplified portion may be used to isolate a full length gene from a suitable library (e.g., human skeletal muscle cell cDNA) using well known techniques. Within such techniques, a library (cDNA or genomic) is screened using one or more polynucleotide probes or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

For hybridization techniques, a partial sequence may be labeled (e.g., by nick-translation or end-labeling with  $^{32}\text{P}$ ) using well known techniques. A bacterial or bacteriophage library may then be screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (see, e.g.,

Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY, 1989). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. Clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a  
5 primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences may be generated to identify one or more overlapping clones. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

Alternatively, there are numerous amplification techniques for obtaining  
10 a full length coding sequence from a partial cDNA sequence. Within such techniques, amplification is generally performed via PCR. One such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, which hybridizes to a polyA region or vector sequence, to identify sequences that are 5' and 3' of a known sequence. Any of a variety of  
15 commercially available kits may be used to perform the amplification step. Primers may be designed using, for example, software well known in the art. Primers are preferably 17-32 nucleotides in length, have a GC content of at least 40% and anneal to the target sequence at temperatures of about 54°C to 72°C. The amplified region may be sequenced as described above, and overlapping sequences assembled into a  
20 contiguous sequence.

A cDNA sequence encoding DSP-15 is provided in Figure 1 (SEQ ID NO:1), and the predicted amino acid sequence is provided in Figure 2 (SEQ ID NO:2). A cDNA sequence encoding a DSP-15 alternate form is provided in Figure 5 (SEQ ID NO:20), and the predicted amino acid sequence is provided in Figure 6 (SEQ ID  
25 NO:21). The DSP-15 active site VHCKMGVSRS (SEQ ID NO:16), is encoded by nucleotide bases located at nucleotide positions 1233 through 1260 of SEQ ID NO:1 (Fig. 1; start codon begins at nucleotide position number 1). Sequence information immediately adjacent to this site was used to design 5' and 3' RACE reactions with human brain, skeletal muscle and testis cDNA to identify a protein of 659 amino acids  
30 encoded by 1977 base pairs. This protein is referred to as dual specificity phosphatase-3, or DSP-15. Higher message abundance was observed for DSP-15 in human skeletal

muscle tissue than in other tissues. DSP-15 shows significant homology to other MAP-kinase phosphatases, as shown by the sequence comparison presented in Figure 3.

DSP-15 (or DSP-15 alternate form) polynucleotide variants may generally be prepared by any method known in the art, including, for example, solid  
5 phase chemical synthesis. Modifications in a polynucleotide sequence may also be introduced using standard mutagenesis techniques, such as oligonucleotide-directed site-specific mutagenesis. Alternatively, RNA molecules may be generated by *in vitro* or *in vivo* transcription of DNA sequences encoding DSP-15, or a portion thereof, provided that the DNA is incorporated into a vector with a suitable RNA polymerase  
10 promoter (such as T7 or SP6). Certain polynucleotides may be used to prepare an encoded polypeptide, as described herein. In addition, or alternatively, a polynucleotide may be administered to a patient such that the encoded polypeptide is generated *in vivo*.

A polynucleotide that is complementary to at least a portion of a coding sequence (*e.g.*, an antisense polynucleotide or a ribozyme) may also be used as a probe  
15 or primer, or to modulate gene expression. Identification of oligonucleotides and ribozymes for use as antisense agents, and DNA encoding genes for their targeted delivery, involve methods well known in the art. For example, the desirable properties, lengths and other characteristics of such oligonucleotides are well known. Antisense oligonucleotides are typically designed to resist degradation by endogenous nucleolytic  
20 enzymes by using such linkages as: phosphorothioate, methylphosphonate, sulfone, sulfate, ketyl, phosphorodithioate, phosphoramidate, phosphate esters, and other such linkages (*see, e.g.*, Agrwal et al., *Tetrahedron Lett.* 28:3539-3542 (1987); Miller et al., *J. Am. Chem. Soc.* 93:6657-6665 (1971); Stec et al., *Tetrahedron Lett.* 26:2191-2194 (1985); Moody et al., *Nucl. Acids Res.* 12:4769-4782 (1989); Uznanski et al., *Nucl.*  
25 *Acids Res.* (1989); Letsinger et al., *Tetrahedron* 40:137-143 (1984); Eckstein, *Annu. Rev. Biochem.* 54:367-402 (1985); Eckstein, *Trends Biol. Sci.* 14:97-100 (1989); Stein In: *Oligodeoxynucleotides. Antisense Inhibitors of Gene Expression*, Cohen, Ed, Macmillan Press, London, pp. 97-117 (1989); Jager et al., *Biochemistry* 27:7237-7246 (1988)).

30 Antisense polynucleotides are oligonucleotides that bind in a sequence-specific manner to nucleic acids, such as mRNA or DNA. When bound to mRNA that

has complementary sequences, antisense prevents translation of the mRNA (*see, e.g.*, U.S. Patent No. 5,168,053 to Altman et al.; U.S. Patent No. 5,190,931 to Inouye, U.S. Patent No. 5,135,917 to Burch; U.S. Patent No. 5,087,617 to Smith and Clusel et al. (1993) *Nucl. Acids Res.* 21:3405-3411, which describes dumbbell antisense oligonucleotides). Triplex molecules refer to single DNA strands that bind duplex DNA forming a colinear triplex molecule, thereby preventing transcription (*see, e.g.*, U.S. Patent No. 5,176,996 to Hogan et al., which describes methods for making synthetic oligonucleotides that bind to target sites on duplex DNA).

Particularly useful antisense nucleotides and triplex molecules are molecules that are complementary to or bind the sense strand of DNA or mRNA that encodes a DSP-15 or a DSP-15 alternate form polypeptide or a protein mediating any other process related to expression of endogenous DSP-15 (or DSP-15 alternate form), such that inhibition of translation of mRNA encoding the DSP-15 (or DSP-15 alternate form) polypeptide is effected. cDNA constructs that can be transcribed into antisense RNA may also be introduced into cells or tissues to facilitate the production of antisense RNA. Antisense technology can be used to control gene expression through interference with binding of polymerases, transcription factors or other regulatory molecules (*see* Gee et al., *In* Huber and Carr, *Molecular and Immunologic Approaches*, Futura Publishing Co. (Mt. Kisco, NY; 1994)). Alternatively, an antisense molecule may be designed to hybridize with a control region of a DSP-15 gene (*e.g.*, promoter, enhancer or transcription initiation site), and block transcription of the gene; or to block translation by inhibiting binding of a transcript to ribosomes.

The present invention also contemplates DSP-15- (or DSP-15 alternate form) specific ribozymes. A ribozyme is an RNA molecule that specifically cleaves RNA substrates, such as mRNA, resulting in specific inhibition or interference with cellular gene expression. There are at least five known classes of ribozymes involved in the cleavage and/or ligation of RNA chains. Ribozymes can be targeted to any RNA transcript and can catalytically cleave such transcripts (*see, e.g.*, U.S. Patent No. 5,272,262; U.S. Patent No. 5,144,019; and U.S. Patent Nos. 5,168,053, 5,180,818, 5,116,742 and 5,093,246 to Cech et al.). Any DSP-15 (or DSP-15 alternate form) mRNA-specific ribozyme, or a nucleic acid encoding such a ribozyme, may be

delivered to a host cell to effect inhibition of DSP-15 gene expression. Ribozymes may therefore be delivered to the host cells by DNA encoding the ribozyme linked to a eukaryotic promoter, such as a eukaryotic viral promoter, such that upon introduction into the nucleus, the ribozyme will be directly transcribed.

5 Any polynucleotide may be further modified to increase stability *in vivo*. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiester linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queosine and wybutosine, as well as acetyl-, methyl-, thio- and  
10 other modified forms of adenine, cytidine, guanine, thymine and uridine.

Nucleotide sequences as described herein may be joined to a variety of other nucleotide sequences using established recombinant DNA techniques. For example, a polynucleotide may be cloned into any of a variety of cloning vectors, including plasmids, phagemids, lambda phage derivatives and cosmids. Vectors of  
15 particular interest include expression vectors, replication vectors, probe generation vectors and sequencing vectors. In general, a suitable vector contains an origin of replication functional in at least one organism, convenient restriction endonuclease sites and one or more selectable markers. Other elements will depend upon the desired use, and will be apparent to those having ordinary skill in the art.

20 Within certain embodiments, polynucleotides may be formulated so as to permit entry into a cell of a mammal, and expression therein. Such formulations are particularly useful for therapeutic purposes, as described below. Those having ordinary skill in the art will appreciate that there are many ways to achieve expression of a polynucleotide in a target cell, and any suitable method may be employed. For  
25 example, a polynucleotide may be incorporated into a viral vector using well known techniques. A viral vector may additionally transfer or incorporate a gene for a selectable marker (to aid in the identification or selection of transduced cells) and/or a targeting moiety, such as a gene that encodes a ligand for a receptor on a specific target cell, to render the vector target specific. Targeting may also be accomplished using an  
30 antibody, by methods known to those having ordinary skill in the art.

Other formulations for therapeutic purposes include colloidal dispersion systems, such as macromolecule complexes, nanocapsules, microspheres, beads, and lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. A preferred colloidal system for use as a delivery vehicle *in vitro* and *in vivo* is a liposome (*i.e.*, an artificial membrane vesicle). The preparation and use of such systems is well known in the art.

Within other aspects, a DSP-15 promoter may be isolated using standard techniques. The present invention provides nucleic acid molecules comprising such a promoter sequence or one or more cis- or trans-acting regulatory elements thereof. Such regulatory elements may enhance or suppress expression of DSP-15 (or DSP-15 alternate form). A 5' flanking region may be generated using standard techniques, based on the genomic sequence provided herein. If necessary, additional 5' sequences may be generated using PCR-based or other standard methods. The 5' region may be subcloned and sequenced using standard methods. Primer extension and/or RNase protection analyses may be used to verify the transcriptional start site deduced from the cDNA.

To define the boundary of the promoter region, putative promoter inserts of varying sizes may be subcloned into a heterologous expression system containing a suitable reporter gene without a promoter or enhancer. Suitable reporter genes may include genes encoding luciferase, beta-galactosidase, chloramphenicol acetyl transferase, secreted alkaline phosphatase or the Green Fluorescent Protein gene. Suitable expression systems are well known and may be prepared using well known techniques or obtained commercially. Internal deletion constructs may be generated using unique internal restriction sites or by partial digestion of non-unique restriction sites. Constructs may then be transfected into cells that display high levels of DSP-15 expression. In general, the construct with the minimal 5' flanking region showing the highest level of expression of reporter gene is identified as the promoter. Such promoter regions may be linked to a reporter gene and used to evaluate agents for the ability to modulate DSP-15 transcription.

Once a functional promoter is identified, cis- and trans-acting elements may be located. Cis-acting sequences may generally be identified based on homology to previously characterized transcriptional motifs. Point mutations may then be

generated within the identified sequences to evaluate the regulatory role of such sequences. Such mutations may be generated using site-specific mutagenesis techniques or a PCR-based strategy. The altered promoter is then cloned into a reporter gene expression vector, as described above, and the effect of the mutation on reporter  
5 gene expression is evaluated.

The present invention also contemplates the use of allelic variants of DSP-15 (or DSP-15 alternate form), as well as DSP-15 sequences from other organisms. Such sequences may generally be identified based upon similarity to the sequences provided herein (*e.g.*, using hybridization techniques) and based upon the  
10 presence of DSP-15 activity, using an assay provided herein.

In general, polypeptides and polynucleotides as described herein are isolated. An "isolated" polypeptide or polynucleotide is one that is removed from its original environment. For example, a naturally-occurring protein is isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably,  
15 such polypeptides are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure. A polynucleotide is considered to be isolated if, for example, it is cloned into a vector that is not a part of the natural environment.

#### ASSAYS FOR DETECTING DSP-15 ACTIVITY

20 According to the present invention, substrates of DSP-15 (or DSP-15 alternate form) may include full length tyrosine phosphorylated proteins and polypeptides as well as fragments (*e.g.*, portions), derivatives or analogs thereof that can be phosphorylated at a tyrosine residue and that may, in certain preferred embodiments, also be able to undergo phosphorylation at a serine or a threonine residue. Such  
25 fragments, derivatives and analogs include any naturally occurring or artificially engineered DSP-15 substrate polypeptide that retains at least the biological function of interacting with a DSP-15 (or DSP-15 alternate form) as provided herein, for example by forming a complex with a DSP-15 (or DSP-15 alternate form). A fragment, derivative or analog of a DSP-15 substrate polypeptide, including substrates that are  
30 fusion proteins, may be (i) one in which one or more of the amino acid residues are

substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue), and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the substrate polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (*e.g.*, polyethylene glycol) or a detectable moiety such as a reporter molecule, or (iv) one in which additional amino acids are fused to the substrate polypeptide, including amino acids that are employed for purification of the substrate polypeptide or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be within the scope of those skilled in the art. In preferred embodiment, a MAP-kinase polypeptide is a substrate for use as provided herein.

DSP-15 (or DSP-15 alternate form) polypeptide variants may be tested for DSP-15 activity using any suitable assay for MAP-kinase phosphatase activity. Such assays may be performed *in vitro* or within a cell-based assay. For example, a MAP-kinase may be obtained in inactive form from Upstate Biotechnology (Lake Placid, NY; catalog number 14-198), for use as a DSP-15 substrate as provided herein. Phosphorylation of the MAP-kinase can be performed using well known techniques (such as those described by Zheng and Guan, *J. Biol. Chem.* 268:16116-16119, 1993) using the MAP-kinase kinase MEK-1 (available from Upstate Biotechnology; cat. no. 14-206).

For example, [<sup>32</sup>P]-radiolabeled substrate (*e.g.*, MAP-kinase) may be used for the kinase reaction, resulting in radiolabeled, activated MAP-kinase. A DSP-15 (or DSP-15 alternate form) polypeptide may then be tested for the ability to dephosphorylate an activated MAP-kinase by contacting the DSP-15 (or DSP-15 alternate form) polypeptide with the MAP-kinase under suitable conditions (*e.g.*, Tris, pH 7.5, 1 mM EDTA, 1 mM dithiothreitol, 1 mg/mL bovine serum albumin for 10 minutes at 30°C; or as described by Zheng and Guan, *J. Biol. Chem.* 268:16116-16119, 1993). Dephosphorylation of the MAP-kinase may be detected using any of a variety of assays, such as a coupled kinase assay (evaluating phosphorylation of a MAP-kinase substrate using any assay generally known in the art) or directly, based on (1) the loss of radioactive phosphate groups (*e.g.*, by gel electrophoresis, followed by



autoradiography); (2) the shift in electrophoretic mobility following dephosphorylation; (3) the loss of reactivity with an antibody specific for phosphotyrosine or phosphothreonine; or (4) a phosphoamino acid analysis of the MAP-kinase. Certain assays may generally be performed as described by Ward et al., *Nature* 367:651-654, 1994 or Alessi et al., *Oncogene* 8:2015-2020, 1993. In general, contact of 500 pg - 50 ng of DSP-15 polypeptide with 100ng - 100  $\mu$ g activated MAP-kinase should result in a detectable dephosphorylation of the MAP-kinase, typically within 20-30 minutes. Within certain embodiments, 0.01 - 10 units/mL (preferably about 0.1 units/mL, where a unit is an amount sufficient to dephosphorylate 1 nmol substrate per minute) DSP-15 polypeptide may be contacted with 0.1 - 10  $\mu$ M (preferably about 1  $\mu$ M) activated MAP-kinase to produce a detectable dephosphorylation of a MAP-kinase. Preferably, a DSP-15 polypeptide results in a dephosphorylation of a MAP-kinase or a phosphorylated substrate (such as a tyrosine- and/or serine-phosphorylated peptide) that is at least as great as the dephosphorylation observed in the presence of a comparable amount of native human DSP-15. It will be apparent that other substrates identified using a substrate trapping mutant as described herein may be substituted for the MAP-kinase within such assays.

#### ANTIBODIES AND ANTIGEN-BINDING FRAGMENTS

Also contemplated by the present invention are peptides, polypeptides, and other non-peptide molecules that specifically bind to a DSP-15 (or DSP-15 alternate form). As used herein, a molecule is said to "specifically bind" to a DSP-15 (or DSP-15 alternate form) if it reacts at a detectable level with DSP-15 (or DSP-15 alternate form), but does not react detectably with peptides containing an unrelated sequence, or a sequence of a different phosphatase. Preferred binding molecules include antibodies, which may be, for example, polyclonal, monoclonal, single chain, chimeric, anti-idiotypic, or CDR-grafted immunoglobulins, or fragments thereof, such as proteolytically generated or recombinantly produced immunoglobulin F(ab')<sub>2</sub>, Fab, Fv, and Fd fragments. Certain preferred antibodies are those antibodies that inhibit or block DSP-15 activity within an *in vitro* assay, as described herein. Binding properties of an antibody to DSP-15 may generally be assessed using immunodetection methods

including, for example, an enzyme-linked immunosorbent assay (ELISA), immunoprecipitation, immunoblotting and the like, which may be readily performed by those having ordinary skill in the art.

Methods well known in the art may be used to generate antibodies, polyclonal antisera or monoclonal antibodies that are specific for a DSP-15 (or DSP-15 alternate form). Antibodies also may be produced as genetically engineered immunoglobulins (Ig) or Ig fragments designed to have desirable properties. For example, by way of illustration and not limitation, antibodies may include a recombinant IgG that is a chimeric fusion protein having at least one variable (V) region domain from a first mammalian species and at least one constant region domain from a second, distinct mammalian species. Most commonly, a chimeric antibody has murine variable region sequences and human constant region sequences. Such a murine/human chimeric immunoglobulin may be "humanized" by grafting the complementarity determining regions (CDRs) derived from a murine antibody, which confer binding specificity for an antigen, into human-derived V region framework regions and human-derived constant regions. Fragments of these molecules may be generated by proteolytic digestion, or optionally, by proteolytic digestion followed by mild reduction of disulfide bonds and alkylation. Alternatively, such fragments may also be generated by recombinant genetic engineering techniques.

As used herein, an antibody is said to be "immunospecific" or to "specifically bind" a DSP-15 (or DSP-15 alternate form) polypeptide if it reacts at a detectable level with DSP-15 (or DSP-15 alternate form), preferably with an affinity constant,  $K_a$ , of greater than or equal to about  $10^4 \text{ M}^{-1}$ , more preferably of greater than or equal to about  $10^5 \text{ M}^{-1}$ , more preferably of greater than or equal to about  $10^6 \text{ M}^{-1}$ , and still more preferably of greater than or equal to about  $10^7 \text{ M}^{-1}$ . Affinities of binding partners or antibodies can be readily determined using conventional techniques, for example, those described by Scatchard et al. (*Ann. N.Y. Acad. Sci. USA* 51:660 (1949)) or by surface plasmon resonance (BIAcore, Biosensor, Piscataway, NJ). See, e.g., Wolff et al., *Cancer Res.* 53:2560-2565 (1993).

Antibodies may generally be prepared by any of a variety of techniques known to those having ordinary skill in the art. See, e.g., Harlow et al., *Antibodies: A*

*Laboratory Manual*, Cold Spring Harbor Laboratory (1988). In one such technique, an animal is immunized with DSP-15 as an antigen to generate polyclonal antisera. Suitable animals include, for example, rabbits, sheep, goats, pigs, cattle, and may also include smaller mammalian species, such as mice, rats, and hamsters, or other species.

5           An immunogen may be comprised of cells expressing DSP-15 (or DSP-15 alternate form), purified or partially purified DSP-15 (or DSP-15 alternate form) polypeptides or variants or fragments (e.g., peptides) thereof, or DSP-15 peptides. DSP-15 peptides may be generated by proteolytic cleavage or may be chemically synthesized. For instance, nucleic acid sequences encoding DSP-15 (or DSP-15  
10 alternate form) polypeptides are provided herein, such that those skilled in the art may routinely prepare these polypeptides for use as immunogens. Polypeptides or peptides useful for immunization may also be selected by analyzing the primary, secondary, and tertiary structure of DSP-15 according to methods known to those skilled in the art, in order to determine amino acid sequences more likely to generate an antigenic response  
15 in a host animal. See, e.g., Novotny, 1991 *Mol. Immunol.* 28:201-207; Berzofsky, 1985 *Science* 229:932-40.

Preparation of the immunogen for injection into animals may include covalent coupling of the DSP-15 (or DSP-15 alternate form) polypeptide (or variant or fragment thereof), to another immunogenic protein, for example, a carrier protein such  
20 as keyhole limpet hemocyanin (KLH) or bovine serum albumin (BSA). In addition, the DSP-15 peptide, polypeptide, or DSP-15-expressing cells to be used as immunogen may be emulsified in an adjuvant. See, e.g., Harlow et al., *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory (1988). In general, after the first injection, animals receive one or more booster immunizations according to a preferred schedule  
25 that may vary according to, *inter alia*, the antigen, the adjuvant (if any) and/or the particular animal species. The immune response may be monitored by periodically bleeding the animal, separating the sera out of the collected blood, and analyzing the sera in an immunoassay, such as an ELISA or Ouchterlony diffusion assay, or the like, to determine the specific antibody titer. Once an antibody titer is established, the  
30 animals may be bled periodically to accumulate the polyclonal antisera. Polyclonal antibodies that bind specifically to the DSP-15 polypeptide or peptide may then be

purified from such antisera, for example, by affinity chromatography using protein A, or the DSP-15 polypeptide, immobilized on a suitable solid support.

Monoclonal antibodies that specifically bind to DSP-15 (or DSP-15 alternate form) polypeptides or fragments or variants thereof, and hybridomas, which  
5 are immortal eukaryotic cell lines, that produce monoclonal antibodies having the desired binding specificity, may also be prepared, for example, using the technique of Kohler and Milstein (*Nature*, 256:495-497; 1976, *Eur. J. Immunol.* 6:511-519 (1975)) and improvements thereto. An animal—for example, a rat, hamster, or preferably mouse—is immunized with a DSP-15 immunogen prepared as described above.  
10 Lymphoid cells that include antibody-forming cells, typically spleen cells, are obtained from an immunized animal and may be immortalized by fusion with a drug-sensitized myeloma (*e.g.*, plasmacytoma) cell fusion partner, preferably one that is syngeneic with the immunized animal and that optionally has other desirable properties (*e.g.*, inability to express endogenous Ig gene products). The lymphoid (*e.g.*, spleen) cells and the  
15 myeloma cells may be combined for a few minutes with a membrane fusion-promoting agent, such as polyethylene glycol or a nonionic detergent, and then plated at low density on a selective medium that supports the growth of hybridoma cells, but not unfused myeloma cells. A preferred selection media is HAT (hypoxanthine, aminopterin, thymidine). After a sufficient time, usually about one to two weeks,  
20 colonies of cells are observed. Single colonies are isolated, and antibodies produced by the cells may be tested for binding activity to the DSP-15 polypeptide, or variant or fragment thereof. Hybridomas producing monoclonal antibodies with high affinity and specificity for a DSP-15 antigen are preferred. Hybridomas that produce monoclonal antibodies that specifically bind to a DSP-15 polypeptide or variant or fragment thereof  
25 are therefore contemplated by the present invention.

Monoclonal antibodies may be isolated from the supernatants of hybridoma cultures. An alternative method for production of a murine monoclonal antibody is to inject the hybridoma cells into the peritoneal cavity of a syngeneic mouse, for example, a mouse that has been treated (*e.g.*, pristane-primed) to promote formation  
30 of ascites fluid containing the monoclonal antibody. Contaminants may be removed from the subsequently (usually within 1-3 weeks) harvested ascites fluid by

conventional techniques, such as chromatography, gel filtration, precipitation, extraction, or the like. For example, antibodies may be purified by affinity chromatography using an appropriate ligand selected based on particular properties of the monoclonal antibody (*e.g.*, heavy or light chain isotype, binding specificity, etc.).

- 5 Examples of a suitable ligand, immobilized on a solid support, include Protein A, Protein G, an anti-constant region (light chain or heavy chain) antibody, an anti-idiotypic antibody and a DSP-15 polypeptide or fragment or variant thereof.

Human monoclonal antibodies may be generated by any number of techniques with which those having ordinary skill in the art will be familiar. Such methods include but are not limited to, Epstein Barr Virus (EBV) transformation of  
10 human peripheral blood cells (*e.g.*, containing B lymphocytes), *in vitro* immunization of human B cells, fusion of spleen cells from immunized transgenic mice carrying human immunoglobulin genes inserted by yeast artificial chromosomes (YAC), isolation from human immunoglobulin V region phage libraries, or other procedures as known in the  
15 art and based on the disclosure herein.

For example, one method for generating human monoclonal antibodies includes immortalizing human peripheral blood cells by EBV transformation. *See, e.g.*, U.S. Patent No. 4,464,456. An immortalized cell line producing a monoclonal antibody that specifically binds to a DSP-15 polypeptide (or a variant or fragment thereof) can be  
20 identified by immunodetection methods as provided herein, for example, an ELISA, and then isolated by standard cloning techniques. Another method to generate human monoclonal antibodies, *in vitro* immunization, includes priming human splenic B cells with antigen, followed by fusion of primed B cells with a heterohybrid fusion partner. *See, e.g.*, Boerner et al., 1991 *J. Immunol.* 147:86-95.

25 Still another method for the generation of human DSP-15-specific monoclonal antibodies and polyclonal antisera for use in the present invention relates to transgenic mice. *See, e.g.*, U.S. Patent No. 5,877,397; Bruggemann et al., 1997 *Curr. Opin. Biotechnol.* 8:455-58; Jakobovits et al., 1995 *Ann. N. Y. Acad. Sci.* 764:525-35. In these mice, human immunoglobulin heavy and light chain genes have been  
30 artificially introduced by genetic engineering in germline configuration, and the endogenous murine immunoglobulin genes have been inactivated. *See, e.g.*,

Bruggemann et al., 1997 *Curr. Opin. Biotechnol.* 8:455-58. For example, human immunoglobulin transgenes may be mini-gene constructs, or transloci on yeast artificial chromosomes, which undergo B cell-specific DNA rearrangement and hypermutation in the mouse lymphoid tissue. See, Bruggemann et al., 1997 *Curr. Opin. Biotechnol.* 8:455-58. Human monoclonal antibodies specifically binding to DSP-15 may be obtained by immunizing the transgenic animals, fusing spleen cells with myeloma cells, selecting and then cloning cells producing antibody, as described above. Polyclonal sera containing human antibodies may also be obtained from the blood of the immunized animals.

10 Chimeric antibodies, specific for a DSP-15, including humanized antibodies, may also be generated according to the present invention. A chimeric antibody has at least one constant region domain derived from a first mammalian species and at least one variable region domain derived from a second, distinct mammalian species. See, e.g., Morrison et al., 1984, *Proc. Natl. Acad. Sci. USA*, 81:6851-55. In preferred embodiments, a chimeric antibody may be constructed by cloning the polynucleotide sequence that encodes at least one variable region domain derived from a non-human monoclonal antibody, such as the variable region derived from a murine, rat, or hamster monoclonal antibody, into a vector containing a nucleic acid sequence that encodes at least one human constant region. See, e.g., Shin et al., 15 1989 *Methods Enzymol.* 178:459-76; Walls et al., 1993 *Nucleic Acids Res.* 21:2921-29. By way of example, the polynucleotide sequence encoding the light chain variable region of a murine monoclonal antibody may be inserted into a vector containing a nucleic acid sequence encoding the human kappa light chain constant region sequence. In a separate vector, the polynucleotide sequence encoding the heavy chain variable region of the monoclonal antibody may be cloned in frame with sequences encoding the human IgG1 constant region. The particular human constant region selected may depend upon the effector functions desired for the particular antibody (e.g., complement fixing, binding to a particular Fc receptor, etc.). Another method known in the art for generating chimeric antibodies is homologous recombination (e.g., U.S. Patent 20 No. 5,482,856). Preferably, the vectors will be transfected into eukaryotic cells for stable expression of the chimeric antibody.

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A non-human/human chimeric antibody may be further genetically engineered to create a "humanized" antibody. Such a humanized antibody may comprise a plurality of CDRs derived from an immunoglobulin of a non-human mammalian species, at least one human variable framework region, and at least one human immunoglobulin constant region. Humanization may in certain embodiments provide an antibody that has decreased binding affinity for a DSP-15 when compared, for example, with either a non-human monoclonal antibody from which a DSP-15 binding variable region is obtained, or a chimeric antibody having such a V region and at least one human C region, as described above. Useful strategies for designing humanized antibodies may therefore include, for example by way of illustration and not limitation, identification of human variable framework regions that are most homologous to the non-human framework regions of the chimeric antibody. Without wishing to be bound by theory, such a strategy may increase the likelihood that the humanized antibody will retain specific binding affinity for a DSP-15, which in some preferred embodiments may be substantially the same affinity for a DSP-15 polypeptide or variant or fragment thereof, and in certain other preferred embodiments may be a greater affinity for DSP-15. *See, e.g., Jones et al., 1986 Nature 321:522-25; Riechmann et al., 1988 Nature 332:323-27.* Designing such a humanized antibody may therefore include determining CDR loop conformations and structural determinants of the non-human variable regions, for example, by computer modeling, and then comparing the CDR loops and determinants to known human CDR loop structures and determinants. *See, e.g., Padlan et al., 1995 FASEB 9:133-39; Chothia et al., 1989 Nature, 342:377-383.* Computer modeling may also be used to compare human structural templates selected by sequence homology with the non-human variable regions. *See, e.g., Bajorath et al., 1995 Ther. Immunol. 2:95-103; EP-0578515-A3.* If humanization of the non-human CDRs results in a decrease in binding affinity, computer modeling may aid in identifying specific amino acid residues that could be changed by site-directed or other mutagenesis techniques to partially, completely or supra-optimally (*i.e.*, increase to a level greater than that of the non-humanized antibody) restore affinity. Those having ordinary skill in the art are familiar with these techniques, and will readily appreciate numerous variations and modifications to such design strategies.

Within certain embodiments, the use of antigen-binding fragments of antibodies may be preferred. Such fragments include Fab fragments or  $F(ab')_2$  fragments, which may be prepared by proteolytic digestion with papain or pepsin, respectively. The antigen binding fragments may be separated from the Fc fragments by  
5 affinity chromatography, for example, using immobilized protein A or protein G, or immobilized DSP-15 polypeptide, or a suitable variant or fragment thereof. Those having ordinary skill in the art can routinely and without undue experimentation determine what is a suitable variant or fragment based on characterization of affinity purified antibodies obtained, for example, using immunodetection methods as provided  
10 herein. An alternative method to generate Fab fragments includes mild reduction of  $F(ab')_2$  fragments followed by alkylation. See, e.g., Weir, *Handbook of Experimental Immunology*, 1986, Blackwell Scientific, Boston.

According to certain embodiments, non-human, human, or humanized heavy chain and light chain variable regions of any of the above described Ig molecules  
15 may be constructed as single chain Fv (sFv) polypeptide fragments (single chain antibodies). See, e.g., Bird et al., 1988 *Science* 242:423-426; Huston et al., 1988 *Proc. Natl. Acad. Sci. USA* 85:5879-5883. Multi-functional sFv fusion proteins may be generated by linking a polynucleotide sequence encoding an sFv polypeptide in-frame with at least one polynucleotide sequence encoding any of a variety of known effector  
20 proteins. These methods are known in the art, and are disclosed, for example, in EP-B1-0318554, U.S. Patent No. 5,132,405, U.S. Patent No. 5,091,513, and U.S. Patent No. 5,476,786. By way of example, effector proteins may include immunoglobulin constant region sequences. See, e.g., Hollenbaugh et al., 1995 *J. Immunol. Methods* 188:1-7. Other examples of effector proteins are enzymes. As a non-limiting example,  
25 such an enzyme may provide a biological activity for therapeutic purposes (see, e.g., Siemers et al., 1997 *Bioconjug. Chem.* 8:510-19), or may provide a detectable activity, such as horseradish peroxidase-catalyzed conversion of any of a number of well-known substrates into a detectable product, for diagnostic uses. Still other examples of sFv fusion proteins include Ig-toxin fusions, or immunotoxins, wherein the sFv polypeptide  
30 is linked to a toxin. Those having ordinary skill in the art will appreciate that a wide variety of polypeptide sequences have been identified that, under appropriate



conditions, are toxic to cells. As used herein, a toxin polypeptide for inclusion in an immunoglobulin-toxin fusion protein may be any polypeptide capable of being introduced to a cell in a manner that compromises cell survival, for example, by directly interfering with a vital function or by inducing apoptosis. Toxins thus may include, for  
5 example, ribosome-inactivating proteins, such as *Pseudomonas aeruginosa* exotoxin A, plant gelonin, bryodin from *Bryonia dioica*, or the like. See, e.g., Thrush et al., 1996 *Annu. Rev. Immunol.* 14:49-71; Frankel et al., 1996 *Cancer Res.* 56:926-32. Numerous other toxins, including chemotherapeutic agents, anti-mitotic agents, antibiotics, inducers of apoptosis (or "apoptogens", see, e.g., Green and Reed, 1998, *Science*  
10 281:1309-1312), or the like, are known to those familiar with the art, and the examples provided herein are intended to be illustrative without limiting the scope and spirit of the invention.

The sFv may, in certain embodiments, be fused to peptide or polypeptide domains that permit detection of specific binding between the fusion protein and  
15 antigen (e.g., a DSP-15). For example, the fusion polypeptide domain may be an affinity tag polypeptide. Binding of the sFv fusion protein to a binding partner (e.g., a DSP-15) may therefore be detected using an affinity polypeptide or peptide tag, such as an avidin, streptavidin or a His (e.g., polyhistidine) tag, by any of a variety of techniques with which those skilled in the art will be familiar. Detection techniques may also  
20 include, for example, binding of an avidin or streptavidin fusion protein to biotin or to a biotin mimetic sequence (see, e.g., Luo et al., 1998 *J. Biotechnol.* 65:225 and references cited therein), direct covalent modification of a fusion protein with a detectable moiety (e.g., a labeling moiety), non-covalent binding of the fusion protein to a specific labeled reporter molecule, enzymatic modification of a detectable substrate by a fusion protein  
25 that includes a portion having enzyme activity, or immobilization (covalent or non-covalent) of the fusion protein on a solid-phase support.

The sFv fusion protein of the present invention, comprising a DSP-15-specific immunoglobulin-derived polypeptide fused to another polypeptide such as an effector peptide having desirable affinity properties, may therefore include, for example,  
30 a fusion protein wherein the effector peptide is an enzyme such as glutathione-S-transferase. As another example, sFv fusion proteins may also comprise a DSP-15-

specific Ig polypeptide fused to a *Staphylococcus aureus* protein A polypeptide; protein A encoding nucleic acids and their use in constructing fusion proteins having affinity for immunoglobulin constant regions are disclosed generally, for example, in U.S. Patent 5,100,788. Other useful affinity polypeptides for construction of sFv fusion proteins may include streptavidin fusion proteins, as disclosed, for example, in WO 89/03422; U.S. 5,489,528; U.S. 5,672,691; WO 93/24631; U.S. 5,168,049; U.S. 5,272,254 and elsewhere, and avidin fusion proteins (see, e.g., EP 511,747). As provided herein, sFv polypeptide sequences may be fused to fusion polypeptide sequences, including effector protein sequences, that may include full length fusion polypeptides and that may alternatively contain variants or fragments thereof.

An additional method for selecting antibodies that specifically bind to a DSP-15 polypeptide or variant or fragment thereof is by phage display. See, e.g.; Winter et al., 1994 *Annul. Rev. Immunol.* 12:433-55; Burton et al., 1994 *Adv. Immunol.* 57:191-280. Human or murine immunoglobulin variable region gene combinatorial libraries may be created in phage vectors that can be screened to select Ig fragments (Fab, Fv, sFv, or multimers thereof) that bind specifically to a DSP-15 polypeptide or variant or fragment thereof. See, e.g., U.S. Patent No. 5,223,409; Huse et al., 1989 *Science* 246:1275-81; Kang et al., 1991 *Proc. Natl. Acad. Sci. USA* 88:4363-66; Hoogenboom et al., 1992 *J. Molec. Biol.* 227:381-388; Schlebusch et al., 1997 *Hybridoma* 16:47-52 and references cited therein. For example, a library containing a plurality of polynucleotide sequences encoding Ig variable region fragments may be inserted into the genome of a filamentous bacteriophage, such as M13 or a variant thereof, in frame with the sequence encoding a phage coat protein, for instance, gene III or gene VIII of M13, to create an M13 fusion protein. A fusion protein may be a fusion of the coat protein with the light chain variable region domain and/or with the heavy chain variable region domain.

According to certain embodiments, immunoglobulin Fab fragments may also be displayed on the phage particle, as follows. Polynucleotide sequences encoding Ig constant region domains may be inserted into the phage genome in frame with a coat protein. The phage coat fusion protein may thus be fused to an Ig light chain or heavy chain fragment (Fd). For example, from a human Ig library, the polynucleotide

sequence encoding the human kappa constant region may be inserted into a vector in frame with the sequence encoding at least one of the phage coat proteins. Additionally or alternatively, the polynucleotide sequence encoding the human IgG1 CH1 domain may be inserted in frame with the sequence encoding at least one other of the phage coat proteins. A plurality of polynucleotide sequences encoding variable region domains (e.g., derived from a DNA library) may then be inserted into the vector in frame with the constant region-coat protein fusions, for expression of Fab fragments fused to a bacteriophage coat protein.

Phage that display an Ig fragment (e.g., an Ig V-region or Fab) that binds to a DSP-15 polypeptide may be selected by mixing the phage library with DSP-15 or a variant or a fragment thereof, or by contacting the phage library with a DSP-15 polypeptide immobilized on a solid matrix under conditions and for a time sufficient to allow binding. Unbound phage are removed by a wash, which typically may be a buffer containing salt (e.g., NaCl) at a low concentration, preferably with less than 100 mM NaCl, more preferably with less than 50 mM NaCl, most preferably with less than 10 mM NaCl, or, alternatively, a buffer containing no salt. Specifically bound phage are then eluted with an NaCl-containing buffer, for example, by increasing the salt concentration in a step-wise manner. Typically, phage that bind the DSP-15 with higher affinity will require higher salt concentrations to be released. Eluted phage may be propagated in an appropriate bacterial host, and generally, successive rounds of DSP-15 binding and elution can be repeated to increase the yield of phage expressing DSP-15 specific immunoglobulin. Combinatorial phage libraries may also be used for humanization of non-human variable regions. See, e.g., Rosok et al., 1996 *J. Biol. Chem.* 271:22611-18; Rader et al., 1998 *Proc. Natl. Acad. Sci. USA* 95:8910-15. The DNA sequence of the inserted immunoglobulin gene in the phage so selected may be determined by standard techniques. See, Sambrook et al., 1989 *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Press. The affinity selected Ig-encoding sequence may then be cloned into another suitable vector for expression of the Ig fragment or, optionally, may be cloned into a vector containing Ig constant regions, for expression of whole immunoglobulin chains.

Phage display techniques may also be used to select polypeptides, peptides or single chain antibodies that bind to DSP-15. For examples of suitable vectors having multicloning sites into which candidate nucleic acid molecules (*e.g.*, DNA) encoding such peptides or antibodies may be inserted, *see, e.g.*, McLafferty et al., 5 *Gene* 128:29-36, 1993; Scott et al., 1990 *Science* 249:386-390; Smith et al., 1993 *Methods Enzymol.* 217:228-257; Fisch et al., 1996, *Proc. Natl. Acad. Sci. USA* 93:7761-66. The inserted DNA molecules may comprise randomly generated sequences, or may encode variants of a known peptide or polypeptide domain that specifically binds to a DSP-15 polypeptide, or variant or fragment thereof, as provided herein. Generally, the 10 nucleic acid insert encodes a peptide of up to 60 amino acids, more preferably a peptide of 3 to 35 amino acids, and still more preferably a peptide of 6 to 20 amino acids. The peptide encoded by the inserted sequence is displayed on the surface of the bacteriophage. Phage expressing a binding domain for a DSP-15 polypeptide may be selected on the basis of specific binding to an immobilized DSP-15 polypeptide as 15 described above. As provided herein, well-known recombinant genetic techniques may be used to construct fusion proteins containing the fragment thereof. For example, a polypeptide may be generated that comprises a tandem array of two or more similar or dissimilar affinity selected DSP-15 binding peptide domains, in order to maximize binding affinity for DSP-15 of the resulting product.

20 In certain other embodiments, the invention contemplates DSP-15 specific antibodies that are multimeric antibody fragments. Useful methodologies are described generally, for example in Hayden et al. 1997, *Curr Opin. Immunol.* 9:201-12; Coloma et al., 1997 *Nat. Biotechnol.* 15:159-63). For example, multimeric antibody fragments may be created by phage techniques to form miniantibodies (U.S. Patent No. 25 5,910 573) or antibodies (Holliger et al., 1997, *Cancer Immunol. Immunother.* 45:128-130). Multimeric fragments may be generated that are multimers of a DSP-15-specific Fv, or that are bispecific antibodies comprising a DSP-15-specific Fv noncovalently associated with a second Fv having a different antigen specificity. *See, e.g.*, Koelemij et al., 1999 *J. Immunother.* 22:514-24. As another example, a multimeric antibody may 30 comprise a bispecific antibody having two single chain antibodies or Fab fragments. According to certain related embodiments, a first Ig fragment may be specific for a first

antigenic determinant on a DSP-15 polypeptide (or variant or fragment thereof), while a second Ig fragment may be specific for a second antigenic determinant of the DSP-15 polypeptide. Alternatively, in certain other related embodiments, a first immunoglobulin fragment may be specific for an antigenic determinant on a DSP-15 polypeptide or variant or fragment thereof, and a second immunoglobulin fragment may be specific for an antigenic determinant on a second, distinct (*i.e.*, non-DSP-15) molecule. Also contemplated are bispecific antibodies that specifically bind DSP-15, wherein at least one antigen-binding domain is present as a fusion protein.

Introducing amino acid mutations into DSP-15-binding immunoglobulin molecules may be useful to increase the specificity or affinity for DSP-15, or to alter an effector function. Immunoglobulins with higher affinity for DSP-15 may be generated by site-directed mutagenesis of particular residues. Computer assisted three-dimensional molecular modeling may be employed to identify the amino acid residues to be changed, in order to improve affinity for the DSP-15 polypeptide. *See, e.g.*, Mountain et al., 1992, *Biotechnol. Genet. Eng. Rev.* 10: 1-142. Alternatively, combinatorial libraries of CDRs may be generated in M13 phage and screened for immunoglobulin fragments with improved affinity. *See, e.g.*, Glaser et al., 1992, *J. Immunol.* 149:3903-3913; Barbas et al., 1994 *Proc. Natl. Acad. Sci. USA* 91:3809-13; U.S. Patent No. 5,792, 456).

Effector functions may also be altered by site-directed mutagenesis. *See, e.g.*, Duncan et al., 1988 *Nature* 332:563-64; Morgan et al., 1995 *Immunology* 86:319-24; Eghtedarzede-Kondri et al., 1997 *Biotechniques* 23:830-34. For example, mutation of the glycosylation site on the Fc portion of the immunoglobulin may alter the ability of the immunoglobulin to fix complement. *See, e.g.*, Wright et al., 1997 *Trends Biotechnol.* 15:26-32. Other mutations in the constant region domains may alter the ability of the immunoglobulin to fix complement, or to effect antibody-dependent cellular cytotoxicity. *See, e.g.*, Duncan et al., 1988 *Nature* 332:563-64; Morgan et al., 1995 *Immunology* 86:319-24; Sensel et al., 1997 *Mol. Immunol.* 34:1019-29.

The nucleic acid molecules encoding an antibody or fragment thereof that specifically binds DSP-15, as described herein, may be propagated and expressed according to any of a variety of well-known procedures for nucleic acid excision,

ligation, transformation and transfection. Thus, in certain embodiments expression of an antibody fragment may be preferred in a prokaryotic host, such as *Escherichia coli* (see, e.g., Pluckthun et al., 1989 *Methods Enzymol.* 178:497-515). In certain other embodiments, expression of the antibody or a fragment thereof may be preferred in a eukaryotic host cell, including yeast (e.g., *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, and *Pichia pastoris*), animal cells (including mammalian cells) or plant cells. Examples of suitable animal cells include, but are not limited to, myeloma, COS, CHO, or hybridoma cells. Examples of plant cells include tobacco, corn, soybean, and rice cells. By methods known to those having ordinary skill in the art and based on the present disclosure, a nucleic acid vector may be designed for expressing foreign sequences in a particular host system, and then polynucleotide sequences encoding the DSP-15 binding antibody (or fragment thereof) may be inserted. The regulatory elements will vary according to the particular host.

A DSP-15-binding immunoglobulin (or fragment thereof) as described herein may contain a detectable moiety or label such as an enzyme, cytotoxic agent or other reporter molecule, including a dye, radionuclide, luminescent group, fluorescent group, or biotin, or the like. The DSP-15-specific immunoglobulin or fragment thereof may be radiolabeled for diagnostic or therapeutic applications. Techniques for radiolabeling of antibodies are known in the art. See, e.g., Adams 1998 *In Vivo* 12:11-21; Hiltunen 1993 *Acta Oncol.* 32:831-9. Therapeutic applications are described in greater detail below and may include use of the DSP-15-binding antibody (or fragment thereof) in conjunction with other therapeutic agents. The antibody or fragment may also be conjugated to a cytotoxic agent as known in the art and provided herein, for example, a toxin, such as a ribosome-inactivating protein, a chemotherapeutic agent, an anti-mitotic agent, an antibiotic or the like.

The invention also contemplates the generation of anti-idiotypic antibodies that recognize an antibody (or antigen-binding fragment thereof) that specifically binds to DSP-15 as provided herein, or a variant or fragment thereof. Anti-idiotypic antibodies may be generated as polyclonal antibodies or as monoclonal antibodies by the methods described herein, using an anti-DSP-15 antibody (or antigen-binding fragment thereof) as immunogen. Anti-idiotypic antibodies or fragments thereof

may also be generated by any of the recombinant genetic engineering methods described above, or by phage display selection. An anti-idiotypic antibody may react with the antigen binding site of the anti-DSP-15 antibody such that binding of the anti-DSP-15 antibody to a DSP-15 polypeptide is competitively inhibited. Alternatively, an anti-idiotypic antibody as provided herein may not competitively inhibit binding of an anti-DSP-15 antibody to a DSP-15 polypeptide.

As provided herein and according to methodologies well known in the art, polyclonal and monoclonal antibodies may be used for the affinity isolation of DSP-15 polypeptides. See, e.g., Hermanson et al., *Immobilized Affinity Ligand Techniques*, Academic Press, Inc. New York, 1992. Briefly, an antibody (or antigen-binding fragment thereof) may be immobilized on a solid support material, which is then contacted with a sample comprising the polypeptide of interest (e.g., a DSP-15). Following separation from the remainder of the sample, the polypeptide is then released from the immobilized antibody.

#### 15 METHODS FOR DETECTING DSP-15 EXPRESSION

Certain aspects of the present invention provide methods that employ antibodies raised against DSP-15 (or DSP-15 alternate form), or hybridizing polynucleotides, for diagnostic and assay purposes. Certain assays involve using an antibody or other agent to detect the presence or absence of DSP-15 (or DSP-15 alternate form), or proteolytic fragments thereof. Alternatively, nucleic acid encoding DSP-15 (or DSP-15 alternate form) may be detected, using standard hybridization and/or PCR techniques. Suitable probes and primers may be designed by those having ordinary skill in the art based on the DSP-15 (or DSP-15 alternate form) cDNA sequence provided herein. Assays may generally be performed using any of a variety of samples obtained from a biological source, such as eukaryotic cells, bacteria, viruses, extracts prepared from such organisms and fluids found within living organisms. Biological samples that may be obtained from a patient include blood samples, biopsy specimens, tissue explants, organ cultures and other tissue or cell preparations. A patient or biological source may be a human or non-human animal, a primary cell culture or culture adapted cell line including but not limited to genetically engineered

cell lines that may contain chromosomally integrated or episomal recombinant nucleic acid sequences, immortalized or immortalizable cell lines, somatic cell hybrid cell lines, differentiated or differentiable cell lines, transformed cell lines and the like. In certain preferred embodiments the patient or biological source is a human, and in certain preferred embodiments the biological source is a non-human animal that is a mammal, for example, a rodent (*e.g.*, mouse, rat, hamster, etc.), an ungulate (*e.g.*, bovine) or a non-human primate. In certain other preferred embodiments of the invention, a patient may be suspected of having or being at risk for having a disease associated with altered cellular signal transduction, or may be known to be free of a risk for or presence of such as disease.

To detect DSP-15 (or DSP-15 alternate form) protein, the reagent is typically an antibody, which may be prepared as described below. There are a variety of assay formats known to those having ordinary skill in the art for using an antibody to detect a polypeptide in a sample. See, *e.g.*, Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. For example, the assay may be performed in a Western blot format, wherein a protein preparation from the biological sample is resolved by gel electrophoresis, transferred to a suitable membrane and allowed to react with the antibody. The presence of the antibody on the membrane may then be detected using a suitable detection reagent, as described below.

In another embodiment, the assay involves the use of antibody immobilized on a solid support to bind to the target DSP-15 (or DSP-15 alternate form) and remove it from the remainder of the sample. The bound DSP-15 may then be detected using a second antibody or reagent that contains a reporter group. Alternatively, a competitive assay may be utilized, in which a DSP-15 (or DSP-15 alternate form) polypeptide is labeled with a reporter group and allowed to bind to the immobilized antibody after incubation of the antibody with the sample. The extent to which components of the sample inhibit the binding of the labeled polypeptide to the antibody is indicative of the reactivity of the sample with the immobilized antibody, and as a result, indicative of the level of DSP-15 (or DSP-15 alternate form) in the sample.

The solid support may be any material known to those having ordinary skill in the art to which the antibody may be attached, such as a test well in a microtiter



plate, a nitrocellulose filter or another suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic such as polystyrene or polyvinylchloride. The antibody may be immobilized on the solid support using a variety of techniques known to those in the art, which are amply described in the patent  
5 and scientific literature.

In certain embodiments, the assay for detection of DSP-15 (or DSP-15 alternate form) in a sample is a two-antibody sandwich assay. This assay may be performed by first contacting an antibody that has been immobilized on a solid support, commonly the well of a microtiter plate, with the biological sample, such that DSP-15  
10 (or DSP-15 alternate form) within the sample is allowed to bind to the immobilized antibody (a 30 minute incubation time at room temperature is generally sufficient). Unbound sample is then removed from the immobilized DSP-15/antibody complexes and a second antibody (containing a reporter group such as an enzyme, dye, radionuclide, luminescent group, fluorescent group or biotin) capable of binding to a  
15 different site on the DSP-15 is added. The amount of second antibody that remains bound to the solid support is then determined using a method appropriate for the specific reporter group. For radioactive groups, scintillation counting or autoradiographic methods are generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups and fluorescent groups. Biotin may be detected  
20 using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme). Enzyme reporter groups may generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of the reaction products. Standards and standard additions may be used to determine the level of DSP-15 in a sample, using well known  
25 techniques.

In a related aspect of the present invention, kits for detecting DSP-15 and DSP-15 phosphatase activity are provided. Such kits may be designed for detecting the level of DSP-15 or nucleic acid encoding DSP-15, or may detect phosphatase activity of DSP-15 in a direct phosphatase assay or a coupled phosphatase assay. In general, the  
30 kits of the present invention comprise one or more containers enclosing elements, such as reagents or buffers, to be used in the assay.

A kit for detecting the level of DSP-15 (or DSP-15 alternate form), or nucleic acid encoding DSP-15 (or DSP-15 alternate form), typically contains a reagent that binds to the DSP-15 protein, DNA or RNA. To detect nucleic acid encoding DSP-15, the reagent may be a nucleic acid probe or a PCR primer. To detect DSP-15 protein, the reagent is typically an antibody. Such kits also contain a reporter group suitable for direct or indirect detection of the reagent (*i.e.*, the reporter group may be covalently bound to the reagent or may be bound to a second molecule, such as Protein A, Protein G, immunoglobulin or lectin, which is itself capable of binding to the reagent). Suitable reporter groups include, but are not limited to, enzymes (*e.g.*, horseradish peroxidase), substrates, cofactors, inhibitors, dyes, radionuclides, luminescent groups, fluorescent groups and biotin. Such reporter groups may be used to directly or indirectly detect binding of the reagent to a sample component using standard methods known to those having ordinary skill in the art.

Kits for detecting DSP-15 activity typically comprise a DSP-15 substrate in combination with a suitable buffer. DSP-15 activity may be specifically detected by performing an immunoprecipitation step with a DSP-15-specific antibody prior to performing a phosphatase assay as described above. Other reagents for use in detecting dephosphorylation of substrate may also be provided.

Within certain diagnostic assays, a proliferative disorder may be detected in a patient or any other biological source organism as provided herein based on the presence of an altered DSP-15 (or DSP-15 alternate form) or an altered level of DSP-15 expression. For example, an antibody may distinguish between a wild-type DSP-15 and an altered DSP-15 having a variation in amino acid sequence. Such a variation may be indicative of the presence of a proliferative disorder, or of susceptibility to such a disorder. Hybridization and amplification techniques may be similarly used to detect modified DSP-15 sequences.

#### METHODS FOR IDENTIFYING MODULATORS OF DSP-15 ACTIVITY

In one aspect of the present invention, DSP-15 (or DSP-15 alternate form) polypeptides may be used to identify agents that modulate DSP-15 activity. Such agents may inhibit or enhance signal transduction via a MAP-kinase cascade, leading to

cell proliferation. An agent that modulates DSP-15 activity may alter (*e.g.*, increase or decrease in a statistically significant manner) expression and/or stability of DSP-15, DSP-15 protein activity and/or the ability of DSP-15 to dephosphorylate a substrate. Agents that may be screened within such assays include, but are not limited to, antibodies and antigen-binding fragments thereof, competing substrates or peptides that represent, for example, a catalytic site or a dual phosphorylation motif, antisense polynucleotides and ribozymes that interfere with transcription and/or translation of DSP-15 and other natural and synthetic molecules, for example small molecule inhibitors, that bind to and inactivate DSP-15.

10           Candidate agents for use in a method of screening for a modulator of DSP-15 according to the present invention may be provided as "libraries" or collections of compounds, compositions or molecules. Such molecules typically include compounds known in the art as "small molecules" and having molecular weights less than  $10^5$  daltons, preferably less than  $10^4$  daltons and still more preferably less than  $10^3$  15 daltons. For example, members of a library of test compounds can be administered to a plurality of samples, each containing at least one DSP-15 (or DSP-15 alternate form) polypeptide as provided herein, and then assayed for their ability to enhance or inhibit DSP-15-mediated dephosphorylation of, or binding to, a substrate. Compounds so identified as capable of influencing DSP-15 function (*e.g.*, phosphotyrosine and/or 20 phosphoserine/threonine dephosphorylation) are valuable for therapeutic and/or diagnostic purposes, since they permit treatment and/or detection of diseases associated with DSP-15 activity. Such compounds are also valuable in research directed to molecular signaling mechanisms that involve DSP-15, and to refinements in the discovery and development of future DSP-15 compounds exhibiting greater specificity.

25           Candidate agents further may be provided as members of a combinatorial library, which preferably includes synthetic agents prepared according to a plurality of predetermined chemical reactions performed in a plurality of reaction vessels. For example, various starting compounds may be prepared employing one or more of solid-phase synthesis, recorded random mix methodologies and recorded reaction split 30 techniques that permit a given constituent to traceably undergo a plurality of permutations and/or combinations of reaction conditions. The resulting products

comprise a library that can be screened followed by iterative selection and synthesis procedures, such as a synthetic combinatorial library of peptides (see *e.g.*, PCT/US91/08694, PCT/US91/04666, which are hereby incorporated by reference in their entireties) or other compositions that may include small molecules as provided  
5 herein (see *e.g.*, PCT/US94/08542, EP 0774464, U.S. 5,798,035, U.S. 5,789,172, U.S. 5,751,629, which are hereby incorporated by reference in their entireties). Those having ordinary skill in the art will appreciate that a diverse assortment of such libraries may be prepared according to established procedures, and tested using DSP-15 according to the present disclosure.

10 In certain embodiments, modulating agents may be identified by combining a candidate agent with a DSP-15 (or DSP-15 alternate form) polypeptide or a polynucleotide encoding such a polypeptide, *in vitro* or *in vivo*, and evaluating the effect of the candidate agent on the DSP-15 phosphatase activity using, for example, a representative assay described herein. An increase or decrease in phosphatase activity  
15 can be measured by performing a representative assay provided herein in the presence and absence of a candidate agent. Briefly, a candidate agent may be included in a mixture of active DSP-15 polypeptide and substrate (*e.g.*, a phosphorylated MAP-kinase), with or without pre-incubation with one or more components of the mixture. In general, a suitable amount of antibody or other agent for use in such an assay ranges  
20 from about 0.01  $\mu$ M to about 100  $\mu$ M. The effect of the agent on DSP-15 activity may then be evaluated by quantifying the loss of phosphate from the substrate, and comparing the loss with that achieved using DSP-15 without the addition of a candidate agent. Alternatively, a coupled kinase assay may be used, in which DSP-15 activity is indirectly measured based on MAP-kinase activity.

25 Alternatively, a polynucleotide comprising a DSP-15 promoter operably linked to a DSP-15 coding region or reporter gene may be used to evaluate the effect of a test compound on DSP-15 transcription. Such assays may be performed in cells that express DSP-15 endogenously (*e.g.*, human or other mammalian skeletal muscle, heart, brain, liver or pancreatic cells) or in cells transfected with an expression vector  
30 comprising a DSP-15 promoter linked to a reporter gene. The effect of a test compound

may then be evaluated by assaying the effect on transcription of DSP-15 or the reporter using, for example, a Northern blot analysis or a suitable reporter activity assay.

DSP-15 activity may also be measured in whole cells transfected with a reporter gene whose expression is dependent upon the activation of an appropriate substrate. For example, appropriate cells (*i.e.*, cells that express DSP-15) may be transfected with a substrate-dependent promoter linked to a reporter gene. In such a system, expression of the reporter gene (which may be readily detected using methods well known to those of ordinary skill in the art) depends upon activation of substrate. Dephosphorylation of substrate may be detected based on a decrease in reporter activity. Candidate modulating agents may be added to such a system, as described above, to evaluate their effect on DSP-15 activity.

The present invention further provides methods for identifying a molecule that interacts with, or binds to, DSP-15 (or DSP-15 alternate form). Such a molecule generally associates with DSP-15 with an affinity constant ( $K_a$ ) of at least  $10^4$ , preferably at least  $10^5$ , more preferably at least  $10^6$ , still more preferably at least  $10^7$  and most preferably at least  $10^8$ . Affinity constants may be determined using well known techniques. Methods for identifying interacting molecules may be used, for example, as initial screens for modulating agents, or to identify factors that are involved in the *in vivo* DSP-15 activity. Techniques for substrate trapping, for example using DSP-15 variants or substrate trapping mutants as described above, are also contemplated according to certain embodiments provided herein. In addition to standard binding assays, there are many other techniques that are well known for identifying interacting molecules, including yeast two-hybrid screens, phage display and affinity techniques. Such techniques may be performed using routine protocols, which are well known to those having ordinary skill in the art (*see, e.g.*, Bartel et al., In *Cellular Interactions in Development: A Practical Approach*, D.A. Harley, ed., Oxford University Press (Oxford, UK), pp. 153-179, 1993). Within these and other techniques, candidate interacting proteins (*e.g.*, putative DSP-15 substrates) may be phosphorylated prior to assaying for the presence of DSP-15-binding or interacting proteins.

Within other aspects, the present invention provides animal models in which an animal either does not express a functional DSP-15 (or DSP-15 alternate

form), or expresses an altered DSP-15. Such animals may be generated using standard homologous recombination strategies. Animal models generated in this manner may be used to study activities of DSP-15 polypeptides and modulating agents *in vivo*.

#### METHODS FOR DEPHOSPHORYLATING A SUBSTRATE

5 In another aspect of the present invention, a DSP-15 (or DSP-15 alternate form) polypeptide may be used for dephosphorylating a substrate of DSP-15 as provided herein. In one embodiment, a substrate may be dephosphorylated *in vitro* by incubating a DSP-15 polypeptide with a substrate in a suitable buffer (*e.g.*, Tris, pH 7.5, 1 mM EDTA, 1 mM dithiothreitol, 1 mg/mL bovine serum albumin) for 10 minutes at  
10 30°C. Any compound that can be dephosphorylated by DSP-15, such as a MAP-kinase, may be used as a substrate. In general, the amounts of the reaction components may range from about 50 pg to about 50 ng of DSP-15 polypeptide and from about 10 ng to about 10 µg of substrate. Dephosphorylated substrate may then be purified, for example, by affinity techniques and/or gel electrophoresis. The extent of substrate  
15 dephosphorylation may generally be monitored by adding [ $\gamma$ -<sup>32</sup>P]labeled substrate to a test aliquot, and evaluating the level of substrate dephosphorylation as described herein.

#### METHODS FOR MODULATING CELLULAR RESPONSES

Modulating agents may be used to modulate, modify or otherwise alter (*e.g.*, increase or decrease) cellular responses such as cell proliferation, differentiation  
20 and survival, in a variety of contexts, both *in vivo* and *in vitro*. In general, to so modulate (*e.g.*, increase or decrease in a statistically significant manner) such a response, a cell is contacted with an agent that modulates DSP-15 activity, under conditions and for a time sufficient to permit modulation of DSP-15 activity. Agents that modulate a cellular response may function in any of a variety of ways. For  
25 example, an agent may modulate a pattern of gene expression (*i.e.*, may enhance or inhibit expression of a family of genes or genes that are expressed in a coordinated fashion). A variety of hybridization and amplification techniques are available for evaluating patterns of gene expression. Alternatively, or in addition, an agent may effect apoptosis or necrosis of the cell, and/or may modulate the functioning of the cell

cycle within the cell. (See, *e.g.*, Ashkenazi et al., 1998 *Science*, 281:1305; Thornberry et al., 1998 *Science* 281:1312; Evan et al., 1998 *Science* 281:1317; Adams et al., 1998 *Science* 281:1322; and references cited therein.)

Cells treated as described above may exhibit standard characteristics of  
5 cells having altered proliferation, differentiation or survival properties. In addition, such cells may (but need not) display alterations in other detectable properties, such as contact inhibition of cell growth, anchorage independent growth or altered intercellular adhesion. Such properties may be readily detected using techniques with which those having ordinary skill in the art will be familiar.

#### 10 THERAPEUTIC METHODS

One or more DSP-15 (or DSP-15 alternate form) polypeptides, modulating agents and/or polynucleotides encoding such polypeptides and/or modulating agents may also be used to modulate DSP-15 activity in a patient. As used  
15 herein, a "patient" may be any mammal, including a human, and may be afflicted with a condition associated with DSP-15 activity or may be free of detectable disease. Accordingly, the treatment may be of an existing disease or may be prophylactic. Conditions associated with DSP-15 activity include any disorder associated with cell proliferation, including cancer, graft-versus-host disease (GVHD), autoimmune  
20 diseases, allergy or other conditions in which immunosuppression may be involved, metabolic diseases, abnormal cell growth or proliferation and cell cycle abnormalities. Certain such disorders involve loss of normal MAP-kinase phosphatase activity, leading to uncontrolled cell growth. DSP-15 polypeptides, and polynucleotides encoding such polypeptides, can be used to ameliorate such disorders. Activators of DSP-15 may also  
be used to treat certain disorders, including Duchenne Muscular Dystrophy.

25 For administration to a patient, one or more polypeptides, polynucleotides and/or modulating agents are generally formulated as a pharmaceutical composition. A pharmaceutical composition may be a sterile aqueous or non-aqueous solution, suspension or emulsion, which additionally comprises a physiologically acceptable carrier (*i.e.*, a non-toxic material that does not interfere with the activity of  
30 the active ingredient). Such compositions may be in the form of a solid, liquid or gas

(aerosol). Alternatively, compositions of the present invention may be formulated as a lyophilizate or compounds may be encapsulated within liposomes using well known technology. Pharmaceutical compositions within the scope of the present invention may also contain other components, which may be biologically active or inactive. Such components include, but are not limited to, buffers (*e.g.*, neutral buffered saline or phosphate buffered saline), carbohydrates (*e.g.*, glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, chelating agents such as EDTA or glutathione, stabilizers, dyes, flavoring agents, and suspending agents and/or preservatives.

Any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of the present invention. Carriers for therapeutic use are well known, and are described, for example, in *Remington's Pharmaceutical Sciences*, Mack Publishing Co. (A.R. Gennaro ed. 1985). In general, the type of carrier is selected based on the mode of administration. Pharmaceutical compositions may be formulated for any appropriate manner of administration, including, for example, topical, oral, nasal, intrathecal, rectal, vaginal, sublingual or parenteral administration, including subcutaneous, intravenous, intramuscular, intrasternal, intracavernous, intrameatal or intraurethral injection or infusion. For parenteral administration, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, kaolin, glycerin, starch dextrins, sodium alginate, carboxymethylcellulose, ethyl cellulose, glucose, sucrose and/or magnesium carbonate, may be employed.

A pharmaceutical composition (*e.g.*, for oral administration or delivery by injection) may be in the form of a liquid (*e.g.*, an elixir, syrup, solution, emulsion or suspension). A liquid pharmaceutical composition may include, for example, one or more of the following: sterile diluents such as water for injection, saline solution, preferably physiological saline, Ringer's solution, isotonic sodium chloride, fixed oils such as synthetic mono or diglycerides which may serve as the solvent or suspending medium, polyethylene glycols, glycerin, propylene glycol or other solvents; antibacterial agents such as benzyl alcohol or methyl paraben; antioxidants such as ascorbic acid or



sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. A parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic. The use of physiological saline is preferred, and an injectable pharmaceutical composition is preferably sterile.

The compositions described herein may be formulated for sustained release (*i.e.*, a formulation such as a capsule or sponge that effects a slow release of compound following administration). Such compositions may generally be prepared using well known technology and administered by, for example, oral, rectal or subcutaneous implantation, or by implantation at the desired target site. Sustained-release formulations may contain an agent dispersed in a carrier matrix and/or contained within a reservoir surrounded by a rate controlling membrane. Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of active component release. The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

For pharmaceutical compositions comprising a polynucleotide encoding a DSP-15 polypeptide and/or modulating agent (such that the polypeptide and/or modulating agent is generated *in situ*), the polynucleotide may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid, and bacterial, viral and mammalian expression systems. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer et al., *Science* 259:1745-1749, 1993 and reviewed by Cohen, *Science* 259:1691-1692, 1993. The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

Within a pharmaceutical composition, a DSP-15 (or DSP-15 alternate form) polypeptide, polynucleotide or modulating agent may be linked to any of a variety of compounds. For example, such an agent may be linked to a targeting moiety (*e.g.*, a

monoclonal or polyclonal antibody, a protein or a liposome) that facilitates the delivery of the agent to the target site. As used herein, a "targeting moiety" may be any substance (such as a compound or cell) that, when linked to an agent enhances the transport of the agent to a target cell or tissue, thereby increasing the local concentration of the agent. Targeting moieties include antibodies or fragments thereof, receptors, ligands and other molecules that bind to cells of, or in the vicinity of, the target tissue. An antibody targeting agent may be an intact (whole) molecule, a fragment thereof, or a functional equivalent thereof. Examples of antibody fragments are F(ab')<sub>2</sub>, -Fab', Fab and F[v] fragments, which may be produced by conventional methods or by genetic or protein engineering. Linkage is generally covalent and may be achieved by, for example, direct condensation or other reactions, or by way of bi- or multi-functional linkers. Targeting moieties may be selected based on the cell(s) or tissue(s) toward which the agent is expected to exert a therapeutic benefit.

Pharmaceutical compositions may be administered in a manner appropriate to the disease to be treated (or prevented). An appropriate dosage and a suitable duration and frequency of administration will be determined by such factors as the condition of the patient, the type and severity of the patient's disease, the particular form of the active ingredient and the method of administration. In general, an appropriate dosage and treatment regimen provides the agent(s) in an amount sufficient to provide therapeutic and/or prophylactic benefit (e.g., an improved clinical outcome, such as more frequent complete or partial remissions, or longer disease-free and/or overall survival). For prophylactic use, a dose should be sufficient to prevent, delay the onset of or diminish the severity of a disease associated with cell proliferation.

Optimal dosages may generally be determined using experimental models and/or clinical trials. In general, the amount of polypeptide present in a dose, produced *in situ* by DNA present in a dose, ranges from about 0.01 µg to about 100 µg per kg of host, typically from about 0.1 µg to about 10 µg. The use of the minimum dosage that is sufficient to provide effective therapy is usually preferred. Patients may generally be monitored for therapeutic or prophylactic effectiveness using assays suitable for the condition being treated or prevented, which will be familiar to those

having ordinary skill in the art. Suitable dose sizes will vary with the size of the patient, but will typically range from about 10 mL to about 500 mL for 10-60 kg animal.

The following Example is offered by way of illustration and not by way of limitation.

5

## EXAMPLES

### EXAMPLE 1

#### CLONING AND SEQUENCING cDNA ENCODING DSP-15

5           This Example illustrates the cloning of a cDNA molecule encoding human DSP-15.

A conserved sequence motif surrounding the active site domain of dual-specificity phosphatases was identified as follows: Dual specificity phosphatases belong to the larger family of protein tyrosine phosphatases (PTPs) that share a  
10 conserved catalytic domain containing a cysteine residue situated N-terminal to a stretch of five variable amino acids followed by an arginine residue (Fauman et al., *Trends In Bioch. Sci.* 21:413-417, 1996). DSPs typically contain a PTP active site motif but lack sequence homology to PTPs in other regions (Jia, *Biochem. and Cell Biol.* 75:17-26, 1997). There is, however, no reported consensus sequence that is conserved among  
15 DSPs, nor is a consensus region apparent from examination of the known DSP sequences such as those referred to above.

To derive a longer consensus DSP amino acid sequence motif that would be useful for the identification of new DSP family members, multiple known human dual-specificity phosphatases sequences were aligned and compared. An alignment of  
20 eight amino acid sequences derived from eight human DSPs having MAP-kinase phosphatase activity yielded a conserved homology region consisting of a 24-amino acid peptide sequence containing the PTP active site signature motif. Thus, a candidate peptide having the sequence:

25           NGRVLVHCQAGISRSGTNILAYLM                   SEQ ID NO:17

was used to search the Expressed Sequence Tag database (Nat. Center for Biol. Information, [www.ncbi.nlm.nih.gov/dbEST](http://www.ncbi.nlm.nih.gov/dbEST)). The search employed an algorithm (tblastn) capable of reverse translation of the candidate peptide with iterations allowing  
30 for genetic code degeneracy within default parameters. The search results identified the

EST AK001790, which was aligned with several known PTPs including VHR and several DSPs (Figure 3) and included a PTP active site motif within a larger domain that was not conserved when compared to the other DSP active site domains, which are associated with functional DSP activity. The translated EST did not contain a substrate  
5 recognition loop, which is a conserved domain of other MAP kinase phosphatases,. The translated EST also did not include a complete coding region of an expressed gene such as a gene encoding a DSP-15 having MAP-kinase phosphatase activity.

The active site amino acid sequence of the DSP-like active site domain encoded by AK001790,

10

VLVHCKMGVSRSAATVLAYAMK      SEQ ID NO:18

was resubmitted to a BLAST (tblastn) search of the GenBank EST database and identified two ESTs having sequence overlaps with AK001790: AW952870 (a human  
15 sequence containing the query domain and having a match of only its first 241 nucleotides with AK001790), and AW326161 (a bovine sequence containing the query domain and having a match of only its last 287 nucleotides with AK001790). When AW952870 was submitted to a BLAST (tblastn) search of the GenBank "month" database, its first 355 nucleotides were found to contain exon sequences encoded in the  
20 human HTGS entry AP001885. Querying the GenBank EST database with AW326161 as a BLAST (tblastn) search sequence identified AW732634 as an additional related human EST, which contained a 284 nucleotide overlap with AW326161. AW732634 also exhibited a 60 nucleotide overlap with AW952870 (described above) and contained exon sequences encoded in the HTGS entry AP001885 (described above).

25                   The derived consensus nucleotide sequence from AW952870 and AW732634 was used to design a 5' RACE primer:

GSP1:

30   5'—TGT CGA TGA AGT CAC GGT ACT GCT GGA GGG—3' SEQ ID NO:19

and the following 3' RACE primers:

SP6:

5'—GCC GCA CTG GAA GGA GAC GCA CCG—3'

SEQ ID NO:22

5

GSP7:

5'—GCG CCA GCT GCA GAT CTA CCA GGG CAT—3'

SEQ ID NO:23

5' and 3' RACE (rapid amplification of cDNA ends) analysis (Frohman  
10 et al., *Proc. Nat. Acad. Sci. USA* 85:8998, 1988; Ohara et al., *Proc. Nat. Acad. Sci. USA*  
86:5673, 1989; Loh et al., *Science* 243:217, 1989) was performed using brain, testis and  
skeletal muscle cDNA templates with 5'/3' RACE kits (Roche Molecular Biochemicals,  
Inc. (formerly Boehringer Mannheim), Indianapolis, IN; Clontech, Palo Alto, CA; Life  
Technologies, Gaithersburg, MD) according to the suppliers' instructions. Sequences of  
15 the RACE products provided the basis for designing additional 5' RACE primers:

GSP2:

5'—CAC TTT CCA CAG CTC AGC ACG GAT CGCC—3'

SEQ ID NO:24

20 GSP2.5:

5'—CGC AGA GAC TCC AGG TCG GCC ATA GCC—3'

SEQ ID NO:25

The 3' RACE reaction reached the stop codon and also revealed a splice  
variant encoding a shorter product. The sequence of the 5' RACE reaction substantially  
25 matched the 5' region sequence of another EST database entry, AK000522, but failed  
by a length of 12 nucleotides to extend all the way to the start codon situated in the 5'  
region of AK000522. An additional oligonucleotide corresponding to a sequence in the  
5' untranslated region of AK000522 was therefore designed for use in additional 5'  
RACE and in PCR reactions:

30

PCR—5':

5'—GGG GTT GAG GGA AGG GGC CGT GC—3' SEQ ID NO:26

PCR and RACE reactions were performed using the PCR-5' primer and the GSP2.5 primer, and the reaction products were sequenced according to standard procedures. A cDNA (Figure 1; SEQ ID NO:1) encoding a protein of 659 amino acids (Figure 2; SEQ ID NO:2) was identified as DSP-15. This sequence has significant homology to other MAP-kinase phosphatases (Figure 6), including DSP-12 and DSP-13 (which are disclosed in U.S. Provisional Application No. 60/179,886 filed February 2, 2000, and which is hereby incorporated by reference). A second cDNA (Figure 4, SEQ ID NO:20) encoding a protein of 471 amino acids (Figure 5, SEQ ID NO:21) was also identified as a DSP-15 alternate form, apparently (and according to non-limiting theory) a truncated form produced by alternate splicing of a DSP-15 encoding transcript. As shown in Figure 6, DSP-15 (and DSP-15 alternate form) exhibit high homology with AB036834, a MAP kinase phosphatase from *Drosophila*. The identified cDNA contains the 1977 base pair coding region, as well as associated 5' and 3' untranslated sequences. The active site domain for DSP-15 was localized to the region encoded by nucleotides beginning at position 1233 through 1260 of SEQ ID NO:1 (Fig. 1; start codon begins at nucleotide position number 1). The MAP kinase phosphatases shown in Figure 6 clearly represent a distinct DSP subfamily. Based on the HTGS database entry AP001885, which was identified as described above, the chromosomal location of the gene encoding DSP-15 was assigned to human chromosome 11q.

## EXAMPLE 2

### DSP-15 EXPRESSION IN HUMAN TISSUES

In this example, a DSP-15 encoding nucleic acid sequence is shown to hybridize to human polyA<sup>+</sup> RNA from various tissue sources. Full length DSP-15 encoding cDNA (SEQ ID NO:1) is <sup>32</sup>P-labeled by the random primer method as described in Ausubel et al. (1998 *Current Protocols in Molecular Biology*, Greene Publ.

Assoc. Inc. & John Wiley & Sons, Inc., Boston, MA) for use as a nucleic acid hybridization probe. The probe is hybridized to blots containing human polyA+ RNA derived from multiple human tissues, normalized for the amount of detectable  $\beta$ -actin mRNA (Cat. No. 7759-1; Clontech, Inc., Palo Alto, CA). Blots undergo  
5 prehybridization for 30 min at 68°C in Express Hyb™ solution (Clontech), and then are hybridized with the labeled probe for 1 hour at 68°C in Express Hyb™ solution. The blots are next washed for 40 min at room temperature in 2X SSC, 0.05% SDS, followed by a second wash for 40 min at 50°C in 0.1X SSC, 0.1% SDS. Blots are exposed to Hyperfilm MP™ autoradiographic film (Amersham Life Sciences, Arlington Hts, IL)  
10 overnight. Results are shown in Figure 4, in which the human tissue sources for the RNAs are as follows: Lane 1, heart; lane 2, brain; lane 3, placenta; lane 4, lung; lane 5, liver; lane 6, skeletal muscle; lane 7, kidney; lane 8, pancreas.

### EXAMPLE 3

#### 15 DSP-15 PHOSPHATASE ACTIVITY

Assays of DSP-15 activity using a tyrosine phosphorylated  $^{32}\text{P}$ -labeled EGF receptor autophosphorylation site peptide as substrate are performed essentially as described (Flint et al., 1993 *EMBO J.* 12:1937-1946; Zhang et al., 1994 *Biochem.* 33:2285-2290). A polynucleotide comprising the DSP-15 coding sequence of SEQ ID  
20 NO:1 is cloned into the pGEX expression vector (Pharmacia, Piscataway, NJ) and expressed in *E. coli* as a DSP-15-glutathione-S-transferase (GST) fusion protein according to the supplier's instructions. Affinity isolation of the DSP-15-GST fusion protein on immobilized glutathione (Pharmacia) following extraction is also conducted as recommended by the supplier. All reagents are from Sigma Chemical Co. (St. Louis,  
25 MO) unless otherwise noted. An aliquot (20  $\mu\text{l}$ ) of ice-cold Assay Buffer (25 mM imidazole (EM Science, Gibbstown, NJ)-pH 7.2, 1 mM EDTA, 2 mM dithiothreitol (DTT, Roche Molecular Biochemicals, Indianapolis, IN), 0.25 mg/ml ovalbumin (Calbiochem-Novabiochem, La Jolla, CA)) is added to wells designated as enzyme negative controls. DSP-15 (SEQ ID NO:2) diluted into ice-cold Assay Buffer from a  
30 50% glycerol stock such that this amount of enzyme would utilize less than 20% of the substrate in the assay, is added, 20  $\mu\text{l}$  per well to all wells except enzyme negative



control wells. The plate is agitated for 20 sec to mix the contents of each well and incubated for 13 min at room temperature. For substrate, the autophosphorylation site from the EGF receptor having the amino acid sequence DADEpYL-NH<sub>2</sub> [SEQ ID NO:27] is prepared as a <sup>32</sup>P-labeled substrate peptide essentially as described (Zhang et al., 1994 *Biochem.* 33:2285; specific activity 11 μCi/nMol), diluted to 0.6 μM in Assay Buffer, and added to all wells in 20 μl aliquots. The plate is again agitated and then incubated an additional 13 minutes, at which time 140 μl of an activated charcoal suspension (25 mg/ml in 0.1 M NaH<sub>2</sub>PO<sub>4</sub>, pH≤5) is added to each well, the contents mixed by vortexing, and the plate is then centrifuged 2400 rpm for three min at room temperature in a tabletop centrifuge (Beckman Instruments, Inc., Fullerton, CA). Aliquots (100 μl) of the supernatant fluid in each well are transferred to a beta-scintillation counting plate (Wallac, Inc., Gaithersburg, MD) and <sup>32</sup>P beta emissions are quantified using a Wallac Microbeta™ plate counter according to the manufacturer's recommendations. After subtracting background counts, correcting for enzyme negative control values and normalizing to control wells DSP-15 specific activity for the EGF receptor peptide substrate is calculated to and expressed as nmole/min/mg, and a K<sub>m</sub> value is determined.

From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the present invention is not limited except as by the appended claims.

## CLAIMS

1. An isolated polypeptide having the sequence of DSP-15 recited in SEQ ID NO:2, or a variant thereof that differs in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of the residues in SEQ ID NO:2, such that the polypeptide retains the ability to dephosphorylate an activated MAP-kinase.
2. An isolated polynucleotide that encodes at least ten consecutive amino acids of a polypeptide having a sequence corresponding to SEQ ID NO:2.
3. An isolated polynucleotide that encodes at least fifteen consecutive amino acids of a polypeptide having a sequence corresponding to SEQ ID NO:2.
4. An expression vector comprising a polynucleotide according to claim 2 or 3.
5. A host cell transformed or transfected with an expression vector according to claim 4.
6. An isolated polynucleotide that encodes a polypeptide according to claim 1.
7. A polynucleotide according to claim 6, comprising the sequence recited in SEQ ID NO:1.
8. An expression vector comprising a polynucleotide according to claim 6.

9. A host cell transformed or transfected with an expression vector according to claim 8.
10. An antisense polynucleotide comprising at least 15 consecutive nucleotides complementary to a polynucleotide according to claim 6.
11. An isolated polynucleotide that detectably hybridizes to the complement of the sequence recited in SEQ ID NO:1 under conditions that include a wash in 0.1X SSC and 0.1% SDS at 50 °C for 15 minutes.
12. An expression vector comprising a polynucleotide according to claim 10 or claim 11.
13. A host cell transformed or transfected with an expression vector according to claim 12.
14. A method of producing a DSP-15 polypeptide, comprising the steps of:
  - (a) culturing a host cell according to claim 9 under conditions that permit expression of the DSP-15 polypeptide; and
  - (b) isolating DSP-15 polypeptide from the host cell culture.
15. An isolated antibody, or antigen binding fragment thereof, that specifically binds to a DSP-15 polypeptide having the sequence of SEQ ID NO:2.
16. An antibody or fragment thereof according to claim 15, wherein the antibody is a monoclonal antibody.
17. A pharmaceutical composition comprising an antibody or fragment thereof according to claim 15 in combination with a physiologically acceptable carrier.

18. A method for detecting DSP-15 expression in a sample, comprising:

- (a) contacting a sample with an antibody or an antigen-binding fragment thereof according to claim 15, under conditions and for a time sufficient to allow formation of an antibody/DSP-15 complex; and
- (b) detecting the level of antibody/DSP-15 complex, and therefrom detecting the presence of DSP-15 in a sample.

19. A method according to claim 18, wherein the antibody is linked to a support material.

20. A method according to claim 18, wherein the antibody is linked to a detectable marker.

21. A method according to claim 18, wherein the sample is a biological sample obtained from a patient.

22. A method for detecting DSP-15 expression in a sample, comprising:

- (a) contacting a sample with an antisense polynucleotide according to claim 10 or claim 11; and
- (b) detecting in the sample an amount of DSP-15 polynucleotide that hybridizes to the antisense polynucleotide, and therefrom detecting DSP-15 expression in the sample.

23. A method according to claim 22, wherein the amount of DSP-15 polynucleotide that hybridizes to the antisense polynucleotide is determined using polymerase chain reaction.

24. A method according to claim 22, wherein the amount of DSP-15 polynucleotide that hybridizes to the antisense polynucleotide is determined using a hybridization assay.

25. A method according to claim 22, wherein the sample comprises an RNA or cDNA preparation.

26. A method for screening for an agent that modulates DSP-15 activity, comprising the steps of:

(a) contacting a candidate agent with a polypeptide according to claim 1, under conditions and for a time sufficient to permit interaction between the polypeptide and candidate agent; and

(b) subsequently evaluating the ability of the polypeptide to dephosphorylate a DSP-15 substrate, relative to a predetermined ability of the polypeptide to dephosphorylate the DSP-15 substrate in the absence of candidate agent; and therefrom identifying an agent that modulates DSP-15 activity.

27. A method according to claim 26, wherein the DSP-15 substrate is a MAP-kinase.

28. A method according to claim 26, wherein the candidate agent is a small molecule.

29. A method according to claim 26, wherein the small molecule is present within a combinatorial library.

30. A method for screening for an agent that modulates DSP-15 activity, comprising the steps of:

(a) contacting a candidate agent with a cell comprising a DSP-15 promoter operably linked to a polynucleotide encoding a detectable transcript or protein,

under conditions and for a time sufficient to permit interaction between the promoter and candidate agent; and

(b) subsequently evaluating the expression of the polynucleotide, relative to a predetermined level of expression in the absence of candidate agent; and therefrom identifying an agent that modulates DSP-15 activity.

31. A method according to claim 30, wherein the polynucleotide encodes a DSP-15 polypeptide.

32. A method according to claim 30, wherein the polynucleotide encodes a reporter protein.

33. A method for modulating a proliferative response in a cell, comprising contacting a cell with an agent that modulates DSP-15 activity.

34. A method for modulating differentiation of a cell, comprising contacting a cell with an agent that modulates DSP-15 activity.

35. A method for modulating survival of a cell, comprising contacting a cell with an agent that modulates DSP-15 activity.

36. A method according to any one of claims 33-35, wherein the agent modulates a pattern of gene expression.

37. A method according to any one of claims 33-35, wherein the cell displays contact inhibition of cell growth.

38. A method according to any one of claims 33-35, wherein the cell displays anchorage independent growth.

39. A method according to any one of claims 33-35, wherein the cell displays an altered intercellular adhesion property.
40. A method according to claim 35, wherein the agent modulates apoptosis.
41. A method according to claim 35, wherein the agent modulates the cell cycle.
42. A method according to claim 32, wherein the cell is present within a patient.
43. A method for treating a patient afflicted with a disorder associated with DSP-15 activity, comprising administering to a patient a therapeutically effective amount of an agent that modulates DSP-15 activity.
44. A method according to claim 43, wherein the disorder is selected from the group consisting of Duchenne muscular dystrophy, cancer, graft-versus-host disease, autoimmune diseases, allergies, metabolic diseases, abnormal cell growth, abnormal cell proliferation and cell cycle abnormalities.
45. A DSP-15 substrate trapping mutant polypeptide that differs from the sequence recited in SEQ ID NO:2 in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of the residues in SEQ ID NO:2, such that the polypeptide binds to a substrate with an affinity that is not substantially diminished relative to DSP-15, and such that the ability of the polypeptide to dephosphorylate a substrate is reduced relative to DSP-15.
46. A substrate trapping mutant polypeptide according to claim 45, wherein the polypeptide contains a substitution at position 382 or position 413 of SEQ ID NO:2.

47. A method for screening a molecule for the ability to interact with DSP-15, comprising the steps of:
- (a) contacting a candidate molecule with a polypeptide according to claim 1 under conditions and for a time sufficient to permit the candidate molecule and polypeptide to interact; and
  - (b) detecting the presence or absence of binding of the candidate molecule to the polypeptide, and therefrom determining whether the candidate molecule interacts with DSP-15.
48. A method according to claim 47, wherein the step of detecting comprises an affinity purification step.
49. A method according to claim 47, wherein the step of detecting comprises a yeast two hybrid screen or a screen of a phage display library.
50. An isolated polypeptide comprising the sequence of DSP-15 alternate form recited in SEQ ID NO:21, or a variant thereof that differs in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of the residues in SEQ ID NO:21, such that the polypeptide retains the ability to dephosphorylate an activated MAP-kinase.
51. An isolated polynucleotide that encodes at least ten consecutive amino acids of a polypeptide having a sequence corresponding to SEQ ID NO:21.
52. An isolated polynucleotide that encodes at least fifteen consecutive amino acids of a polypeptide having a sequence corresponding to SEQ ID NO:21.
53. An expression vector comprising a polynucleotide according to claim 51 or 52.



54. A host cell transformed or transfected with an expression vector according to claim 53.
55. An isolated polynucleotide that encodes a polypeptide according to claim 50.
56. A polynucleotide according to claim 55, comprising the sequence recited in SEQ ID NO:20.
57. An expression vector comprising a polynucleotide according to claim 55.
58. A host cell transformed or transfected with an expression vector according to claim 57.
59. An antisense polynucleotide comprising at least 15 consecutive nucleotides complementary to a polynucleotide according to claim 55.
60. An isolated polynucleotide that detectably hybridizes to the complement of the sequence recited in SEQ ID NO:20 under conditions that include a wash in 0.1X SSC and 0.1% SDS at 60 °C for 15 minutes.
61. An expression vector comprising a polynucleotide according to claim 59 or claim 60.
62. A host cell transformed or transfected with an expression vector according to claim 61.
63. A method of producing a DSP-15 alternate form polypeptide, comprising the steps of:

(a) culturing a host cell according to claim 58 under conditions that permit expression of the DSP-15 alternate form polypeptide; and

(b) isolating DSP-15 alternate form polypeptide from the host cell culture.

64. An isolated antibody, or antigen binding fragment thereof, that specifically binds to a DSP-15 alternate form polypeptide having the sequence of SEQ ID NO:21.

65. An antibody or fragment thereof according to claim 64, wherein the antibody is a monoclonal antibody.

66. A pharmaceutical composition comprising an antibody or fragment thereof according to claim 64 in combination with a physiologically acceptable carrier.

67. A method for detecting DSP-15 alternate form expression in a sample, comprising:

(a) contacting a sample with an antibody or an antigen-binding fragment thereof according to claim 64, under conditions and for a time sufficient to allow formation of an antibody/DSP-15 alternate form complex; and

(b) detecting the level of antibody/DSP-15 alternate form complex, and therefrom detecting the presence of DSP-15 alternate form in a sample.

68. A method according to claim 67, wherein the antibody is linked to a support material.

69. A method according to claim 67, wherein the antibody is linked to a detectable marker.

70. A method according to claim 67, wherein the sample is a biological sample obtained from a patient.

71. A method for detecting DSP-15 alternate form expression in a sample, comprising:

(a) contacting a sample with an antisense polynucleotide according to claim 59 or claim 60; and

(b) detecting in the sample an amount of DSP-15 alternate form polynucleotide that hybridizes to the antisense polynucleotide, and therefrom detecting DSP-15 alternate form expression in the sample.

72. A method according to claim 71, wherein the amount of DSP-15 alternate form polynucleotide that hybridizes to the antisense polynucleotide is determined using polymerase chain reaction.

73. A method according to claim 71, wherein the amount of DSP-15 alternate form polynucleotide that hybridizes to the antisense polynucleotide is determined using a hybridization assay.

74. A method according to claim 71, wherein the sample comprises an RNA or cDNA preparation.

75. A method for screening for an agent that modulates DSP-15 alternate form activity, comprising the steps of:

(a) contacting a candidate agent with a polypeptide according to claim 50, under conditions and for a time sufficient to permit interaction between the polypeptide and candidate agent; and

(b) subsequently evaluating the ability of the polypeptide to dephosphorylate a DSP-15 alternate form substrate, relative to a predetermined ability of the polypeptide to dephosphorylate the DSP-15 alternate form substrate in the absence of candidate agent;

and therefrom identifying an agent that modulates DSP-15 alternate form activity.

76. A method according to claim 75, wherein the DSP-15 alternate form substrate is a MAP-kinase.

77. A method according to claim 75, wherein the candidate agent is a small molecule.

78. A method according to claim 75, wherein the small molecule is present within a combinatorial library.

79. A method for screening for an agent that modulates DSP-15 alternate form activity, comprising the steps of:

(a) contacting a candidate agent with a cell comprising a DSP-15 alternate form promoter operably linked to a polynucleotide encoding a detectable transcript or protein, under conditions and for a time sufficient to permit interaction between the promoter and candidate agent; and

(b) subsequently evaluating the expression of the polynucleotide, relative to a predetermined level of expression in the absence of candidate agent; and therefrom identifying an agent that modulates DSP-15 alternate form activity.

80. A method according to claim 79, wherein the polynucleotide encodes a DSP-15 alternate form polypeptide.

81. A method according to claim 79, wherein the polynucleotide encodes a reporter protein.

82. A method for modulating a proliferative response in a cell, comprising contacting a cell with an agent that modulates DSP-15 alternate form activity.

83. A method for modulating differentiation of a cell, comprising contacting a cell with an agent that modulates DSP-15 alternate form activity.

84. A method for modulating survival of a cell, comprising contacting a cell with an agent that modulates DSP-15 alternate form activity.

85. A method according to any one of claims 82-84, wherein the agent modulates a pattern of gene expression.

86. A method according to any one of claims 82-84, wherein the cell displays contact inhibition of cell growth.

87. A method according to any one of claims 82-84, wherein the cell displays anchorage independent growth.

88. A method according to any one of claims 82-84, wherein the cell displays an altered intercellular adhesion property.

89. A method according to claim 84, wherein the agent modulates apoptosis.

90. A method according to claim 84, wherein the agent modulates the cell cycle.

91. A method according to claim 81, wherein the cell is present within a patient.

92. A method for treating a patient afflicted with a disorder associated with DSP-15 alternate form activity, comprising administering to a patient a therapeutically effective amount of an agent that modulates DSP-15 alternate form activity.

93. A method according to claim 92, wherein the disorder is selected from the group consisting of cancer, graft-versus-host disease, autoimmune diseases, allergies, metabolic diseases, abnormal cell growth, abnormal cell proliferation and cell cycle abnormalities.

94. A DSP-15 alternate form substrate trapping mutant polypeptide that differs from the sequence recited in SEQ ID NO:21 in one or more amino acid deletions, additions, insertions or substitutions at no more than 50% of the residues in SEQ ID NO:21, such that the polypeptide binds to a substrate with an affinity that is not substantially diminished relative to DSP-15 alternate form, and such that the ability of the polypeptide to dephosphorylate a substrate is reduced relative to DSP-15 alternate form.

95. A substrate trapping mutant polypeptide according to claim 94, wherein the polypeptide contains a substitution at position 382 or position 413 of SEQ ID NO:21.

96. A method for screening a molecule for the ability to interact with DSP-15 alternate form, comprising the steps of:

(a) contacting a candidate molecule with a polypeptide according to claim 50 under conditions and for a time sufficient to permit the candidate molecule and polypeptide to interact; and

(b) detecting the presence or absence of binding of the candidate molecule to the polypeptide, and therefrom determining whether the candidate molecule interacts with DSP-15 alternate form.

97. A method according to claim 96, wherein the step of detecting comprises an affinity purification step.

98. A method according to claim 96, wherein the step of detecting comprises a yeast two hybrid screen or a screen of a phage display library.

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1 CCGGTGCCAG CCCAGGTGCT CGCGGCCTGG CTCCATGGCC CTGGTCACAG TGAGCCGTTT  
61 GCGCCCGGGC AGCGGCGCCT CCACGCCCGT GGGGCCCTGG GACCAGGCGG TCCAGCGAAG  
121 GAGTCGACTC CAGCGAAGGC AGAGCTTTGC GGTGCTCCGT GGGGCTGTCC TGGGACTGCA  
181 GGATGGAGGG GACAATGATG ATGCAGCAGA GGCCAGTTCT GAGCCAACAG AGAAGGCCCC  
241 GAGTGAGGAG GAGCTCCACG GGGACCAGAC AGACTTCGGG CAAGGATCCC AGAGTCCCCA  
301 GAAGCAGGAG GAGCAGAGGC AGCACCTGCA CCTCATGGTA CAGTGTCTGA GGCCGAGGA  
361 TGACATCCGC CTGGCAGCCC AGCTGGAGGC ACCCGGCCT CCCGGCTCC GCTACCTGCT  
421 GGTAGTTTCT ACACGAGAAG GAGAAGGTCT GAGCCAGGAT GAGACGGTCC TCCTGGGCGT  
481 GGATTTCCCT GACAGCAGCT CCCCAGCTG CACCCTGGGC CTGGTCTTGC CCCTCTGGAG  
541 TGACACCCAG GTGACTTAG ATGGAGACGG GGGCTTCAGC GTGACGTCTG GTGGGCAAG  
601 CCGGATCTTC AAGCCCATCT CCATCCAGAC CATGTGGGCC AACTCCAGG TATTGCACCA  
661 AGCATGTGAG GCAGCTCTAG GCAGCGGCCT TGTACCGGT GGCAGTCCCC TCACCTGGGC  
721 CAGCCACTAC CAGGAGAGAC TGAACCTCGA ACAGAGCTGC CTAAATGAGT GGACGGCTAT  
781 GGCCGACCTG GAGTCTCTGC GGCCTCCAG CGCCGAGCCT GCGGGTCTT CAGAACAGGA  
841 GCAGATGGAG CAGGCGATCC GTGCTGAGCT GTGAAAGTG TTGGATGTCA GTGACCTGGA  
901 GAGTGTCACT TCCAAAGAGA TCCGCCAGGC TCTGGAGCTG CGCTGGGGC TCCCCCTCCA  
961 GCAGTACCGT GACTTCATCG ACAACCAGAT GCTGTGCTG GTGGCACAGC GGGACCGAGC  
1021 CTCCCGCATC TTCCCCACC TCTACCTGGG CTCAGAGTGG AACGCAGCAA ACCTGGAGGA  
1081 GCTGCAGAGG AACAGGGTCA CCCACATCTT GAACATGGCC CGGAGATTG ACAACTTCTA  
1141 CCCTGAGCGC TTCACCTACC ACAATGTGCG CCTCTGGGAT GAGGAGTCGG CCCAGCTGCT  
1201 GCCGCACTGG AAGGAGACGC ACCGCTTCAT TGAGGCTGCA AGAGCACAGG GCACCCACGT  
1261 GCTGGTCCAC TGCAAGATGG GCGTCAGCG CTCAGCGGCC ACAGTCTGG CCTATGCCAT  
1321 GAAGCAGTAC GAATGCAGCC TGGAGCAGGC CCTGCGCCAC GTGCAGGAGC TCCGGCCCAT  
1381 CGCCGCCCC AACCTGGCT TCCTGCGCCA GCTGCAGATC TACCAGGGCA TCCTGACGGC  
1441 CAGCCGCCAG AGCCATGTCT GGGAGCAGAA AGTGGGTGGG GTCTCCCCAG AGGAGCACCC  
1501 AGCCCTGAA GTCTCTACAC CATTCCCACC TCTTCCGCA GAACCTGAGG GTGGTGGGA  
1561 GGAGAAGGTT GTAGGCATGG AAGAGAGCCA GGCAGCCCCG AAAGAAGAGC CTGGGCCACG  
1621 GCCACGTATA AACCTCCGAG GGGTCATGAG GTCCATCAGT CTCTGGAGC CCTCCTTGA  
1681 GCTGGAGAGC ACCTCAGAGA CCAGTGACAT GCCAGAGGTC TTCTCTTCCC ACGAGTCTTC  
1741 ACATGAAGAG CCTCTGCAGC CCTTCCACA GCTTGCAAGG ACCAAGGGAG GCCAGCAGT  
1801 GGACAGGGGG CCTCAGCCTG CCTGAAGTC CCGCCAGTCA GTGGTTACCC TCCAGGGCAG  
1861 TGCCGTGGTG GCCAACCAGA CCCAGGCCTT CCAGGAGCAG GAGCAGGGG AGGGGCAGGG  
1921 GCAGGGAGAG CCCTGCATTT CCTCTACGCC CAGGTTCCGG AAGGTGGTGA GACAGGCAG  
1981 CGTGCAATGAG AGTGGAGAGG AAGGCGAGGC CTGAGCCCTC ACACATGCCC ACGCTCCCT  
2041 GACACTGAAG AGGATCCACA ACTCCTTGA GAAACACCCT CACGTCTGTT GCCGCACACA  
2101 TTCCTCTCAG CTCCGCCCCA TACCGTCCAG TACAGCCTCA CCTCCCACCC CTGTCACTAC  
2161 GGCCTCACCT CCCACCCCTG TCACTACAGC CTCACCTCCT ACAGCCTTAA GTCCAGGGC  
2221 CATGTCTGCC TGTCCAAGGG CTCAAGACTT TCTAACTGGG ATGTGGTAGA GGGACTGAAG  
2281 GTACCTTTGG GGGCAACAGC ACCCTAGTTT CATTCTAAC TCTAGCCCTG CACACTCACC  
2341 TGTGGCACGG AATGAAAACA GAGCTTCCCG TGCAAAAAGG GTCACGCCTC CCACCCCGC

*Fig. 1A*

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2401 CCCCTCCCTG CACCTCCTGT CCTCTCCCAG TTCATTCTG GAACCAGCCA GGCCAGGCAA  
2461 CCAGTGGCCC CCAAAGGCAG GCAGGATCCT CAGGCCCCAG CCGCGGGAGG CTGGAAGGGC  
2521 TGGCAGATCG CTTCCCTCAT CCACCTCCAC CGGTCCAGGT CTTTGCTGCT GTCCCAGAC  
2581 CTCCTGTGAC ACCACGCCAG ATCACAGGGC ACCAGGCCAG AGATAGTCTT CTTTTGTCC  
2641 TTTCTGGCCT CTGGCTAGTC AGTTTTTCAT AGCCTTACAG TATCTGGCTT TGTACTGAGA  
2701 AATAAACAC ATTTTCAT

*Fig. 1B*

MALVTVSRSPPGSGASTPVGPWDQAVQRRSRLQRRQSFVLRGAVLGLQDGGDNDAAEASSEPTKAPSEELHGD  
QTDFGQGSQSPQKQEEQRQHLLHLMVQLLRPQDDIRLAAQLEAPRPPRLRYLLVVSTREGEGLSQDET VLLGVDFPDS  
SSPSC TLGLVPLWSDTQVYLDGDDGFSVTSGGQSRIFKPI SIQTMMATLQVLHQACEAALGSLVPGG SALTWASH  
YQERLNSEQSCLNEW TAMADLES LRPPSAEPGGSSEQEQMEQAI RAELWKVLDVSDLESVTSKEIRQALELRGLPL  
QQYRDFIDNQMLLLVAQRDRASRIFPHLYLGSEWNAANLEELQRNRVTHILNMAREIDNFYPERFTYHNVRLWDEES  
AQLLPHWKETHRFIEAARAQGTHVLVHCKMGVSRSAATV LAYAMKQYEC SLEQALRHVQELRPIARP NPGFLRQLQI  
YQGILTASRQSHVWEQKVGVSPEEHPAPEVSTPFPLPPEPEGGGEKVVGMESQAAPKEEPGRPRINLRGVMR  
SISLLEPSLELESTSETSDMPEVFSSHESSHEEPLQFPQLARTKGGQVDRGPQALKSRSVVTLQGS AVVANRT  
QAFQEQEQGGGQGEPCISSTPRFRKVV RQASVHDSGEEGEA

*Fig. 2*

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1.....10.....20.....30.....40.....50.....60.....70.....80
PYST1 DGSPLNSQSPFVEILPFLYLGCADSTNLDVLEEFGIKYLNVTP-----NLPNLFENAGEFKYKQIPISDHWSON
MKP-7 DGSVPSSQPAFPVQILPYLYLGCADSTNLDVLGKYGIKYLNVTP-----NLPNAFEHGGEFTYKQIPISDHWSON
MKP-4 ATPPVGLRASFPVQILPNLYLGSARDSANLESKLGRILNVTP-----NLPNFFEKNGDFHYKQIPISDHWSON
hVH5 LSPCLPVPVSVGLTRILPHLYLGSQKDVLNKDLMTQNGISVVLNASN-----SCP KP-DFICESRFMRVPINDNYCEK
PAC1 SDPRVIYDQGGPVEILPYLYLGSCHSSDLQGLQACGITAVLNVSA-----SCP NHFE--GLFHYKSIPIVEDNQMV E
MKP-1 SSCSTPLYDQGGPVEILPFLYLSAYHASRKM L D A LGIT AL IN VSA-----NCP NHFE--GHYQYKSIPIVEDNHKAD
MKP-2 SSCGTPLHDQGGPVEILPFLYLSAYHAARDMLDALGITALLNVSS-----DCPNHFE--GHYQYKCIPIVEDNHKAD
MKP-5 NVSYRPAYDQGGPVEILPFLYLSAYHASKCEFLANLHITALLNVSR-----RTSEACM--T L L H Y K W I P V E D S H T A D
VHR DSGGCSLPSQPCNEVTPRIYVGNASVAQDIPKLQKLGITHVLNAAEGRSFMHVTNANFYKOSGITVLGIKANDTQEFN
AK001790 MEGTMMMQRPVLSQQHPFSILNSSPAHSPHAREIDNFYP--ERFTYHNVRWLWDEESAQ
Consensus pveilp.lylgsa.....$....gi...Lnvs.....pn.f.....%k.!p.D.....

81.....90.....100.....110.....120.....130.....140.....150.....160 164
PYST1 LSQFFPEAISFIDEA-RGKNCGVLVHCLAGISRSVTVTVAYLMQKLNLSMNDAYDIVKMKKSNISPNFNFMGQLLDFERTLGLS
MKP-7 LSQFFPEAISFIDEA-RSKKGGVLVHCLAGISRSVTVTVAYLMQKMNLSLNDAYDFVKRKKSNISPNFNFMGQLLDFERTLGLS
MKP-4 LSRFFPEAIEFIDEA-LSQNCGLVHCLAGVRSVTVTVAYLMQKLHLSLNDAYDLVKRKKSNISPNFNFMGQLLDFERSLRLE
hVH5 LLPWLDKSIIEFIDKA-KLSSCQIVHCLAGISRSATIAIAYIMKTGMSSDDAYRFVKDRRPSISPNFNFGLQLLEYERTLKLL
PAC1 ISAWFQEAISFIDSV-KNSGGRVLVHCQAGISRSATICLAYLIQSHRVRLDEAFDFVKORRGVISP NF SFGQLLQLETQVLCH
MKP-1 ISSWFNEAIDFIDSI-KNAGGRV FVHCQAGISRSATICLAYLMRTNRVKLDEAFEFVKORRSIISP NF SFGQLLQFESQVLAP
MKP-2 ISSWFMEAIEYIDAV-KDCRGRVLVHCQAGISRSATICLAYLMKKRVRL EEA FE FVKORRSIISP NF SFGQLLQFESQVLAT
MKP-5 ISSHFQEAIDFIDCV-REKGGKVLVHCEAGISRSPTICMAYLMKTKQFRLKEAFDYIKORSMVSPNFSGFMGQLLQY ESEILPS
VHR LSAYFERAADFIDQALAQKNGRVLVHCREGYSRPTLVIA YLMRQKMDVKLSALSIVRQNR E-IGPNDGFLAQLCQLNDR LAKE
AK001790 LLPHWKETHRFIEAA-RAQGTHVLVHCKMGVSRSAATVLA YAMKQYECSEALRHVQELRPIARPNGFLRQLQIYQGILTAR
Consensus ls..f.eai.FI#.a.....V1VHC.agiRSat...Ay1M.....s1..A....!k..r..isPNF.F$gQL1..#..1....

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Fig. 3

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1 CCGGTGCCAG CCCAGGTGCT CGCGGCTGG CTCCATGGCC CTGGTCACAG TGAGCCGTTC  
 61 GCGCCCGGGC AGCGGCGCCT CCACGCCCGT GGGGCCCTGG GACCAGGCGG TCCAGCGAAG  
 121 GAGTCGACTC CAGCGAAGGC AGAGCTTTGC GGTGCTCCGT GGGGCTGTCC TGGGACTGCA  
 181 GGATGGAGGG GACAATGATG ATGCAGCAGA GGCCAGTTCT GAGCCAACAG AGAAGGCCCC  
 241 GAGTGAGGAG GAGCTCCACG GGGACCAGAC AGACTTCGGG CAAGGATCCC AGAGTCCCCA  
 301 GAAGCAGGAG GAGCAGAGGC AGCACCTGCA CCTCATGGTA CAGCTGCTGA GGCCGCAGGA  
 361 TGACATCCGC CTGGCAGCCC AGCTGGAGGC ACCCGGGCCT CCCCGGCTCC GCTACCTGCT  
 421 GGTAGTTTCT ACACGAGAAG GAGAAGGTCT GAGCCAGGAT GAGACGGTCC TCCTGGGCGT  
 481 GGATTTCCCT GACAGCAGCT CCCCAGCTG CACCCTGGGC CTGGTCTTGC CCCTCTGGAG  
 541 TGACACCCAG GTGTACTTAG ATGGAGACGG GGGCTTCAGC GTGACGTCTG GTGGGCAAG  
 601 CCGGATCTTC AAGCCCATCT CCATCCAGAC CATGTGGGCC AACTCCAGG TATTGCACCA  
 661 AGCATGTGAG GCAGCTCTAG GCAGCGGCCT TGTACCGGT GGCAGTGCCC TCACCTGGGC  
 721 CAGCCACTAC CAGGAGAGAC TGAATCCGA ACAGAGCTGC CTCAATGAGT GGACGGCTAT  
 781 GGCCGACCTG GAGTCTCTGC GGCCTCCAG CGCCGAGCCT GGCGGTCTCT CAGAACAGGA  
 841 GCAGATGGAG CAGGCATCC GTGCTGAGCT GTGGAAAGTG TTGGATGTCA GTGACCTGGA  
 901 GAGTGTCACT TCAAAGAGA TCCGCCAGGC TCTGGAGCTG CGCCTGGGGC TCCCCTCCA  
 961 GCAGTACCGT GACTTCATCG ACAACCAGAT GCTGCTGCTG GTGGCACAGC GGGACCGAGC  
 1021 CTCCCGCATC TTCCCCACC TCTACCTGGG CTCAGAGTGG AACGCAGCAA ACCTGGAGGA  
 1081 GCTGCAGAGG AACAGGTGCA CCCACATCTT GAACATGGCC CGGGAGATTG ACAACTTCTA  
 1141 CCCTGAGCGC TTCACCTACC ACAATGTGCG CCTCTGGGAT GAGGAGTCGG CCCAGCTGCT  
 1201 GCCGCACTGG AAGGAGACGC ACCGCTTCAT TGAGGCTGCA AGAGCACAGG GCACCCACGT  
 1261 GCTGGTCCAC TGCAAGATGG GCGTCAGCCG CTCAGCGGCC ACAGTGCTGG CCTATGCCAT  
 1321 GAAGCAGTAC GAATGCAGCC TGGAGCAGGC CCTGCGCCAC GTGCAGGAGC TCCGGCCCAT  
 1381 CGCCCGCCCC AACCTGGCT TCCTGCGCCA GCTGCAGATC TACCAGGGCA TCCTGACGGC  
 1441 CAGAACCTGA GGGTGGTGGG GAGGAGAAGG TTGTAGGCAT GGAAGAGAGC CAGGCAGCCC  
 1501 CGAAAGAAGA GCCTGGGCCA CGGCCACGTA TAAACCTCCG AGGGGTGATG AGGTCCATCA  
 1561 GTCTTCTGGA GCCCTCCTTG GAGCTGGAGA GCACCTCAGA GACCAGTGAC ATGCCAGAGG  
 1621 TCTTCTCTTC CCACGAGTCT TCACATGAAG AGCCTCTGCA GCCCTTCCA CAGCTTGCAA  
 1681 GGACCAAGGG AGGCCAGCAG GTGGACAGGG GGCCTCAGCC TGCCCTGAAG TCCCGCCAGT  
 1741 CAGTGGTTAC CCTCCAGGGC AGTGCCGTGG TGCCCAACCG GACCCAGGCC TTCCAGGAGC  
 1801 AGGAGCAGGG GCAGGGGAG GGGCAGGGAG AGCCTGTCAT TTCCTCTACG CCCAGGTTCC  
 1861 GGAAGGTGGT GAGACAGGCC AGCGTGCATG ACAGTGGAGA GGAGGGCGAG GCCTGAGCCC  
 1921 TCACACATGC CCACGCTCCC CTGACACTGA AGAGGATCCA CAACTCCTTG GAGAAACACC  
 1981 CTCACGTCTG TTGCCGCACA CATTCTCTC AGCTCCGGCC CATACCCGTC ACTACAGCCT  
 2041 CACCTCCCAC CCCTGTCACT ACGGCCTCAC CTCCCACCCC TGTACTACA GCCTCACCTC  
 2101 CTACAGCCTT AAGTCCCAGG CCCATGTCTG CCTGTCCAAG GGCTCAAGAC TTTCTAATG  
 2161 GGATGTGGTA GAGGGACTGA AGGTACCTTT GGGGGCAACA GCACCCTAGT TTCATTCTCA  
 2221 ACTCTAGCCC TGCACACTCA CCTGTGGCAC GGAATGAAAA CAGAGCTTCC CGTGCAAAAA  
 2281 GGGTCACGCC TCCCACCCC GCCCCCTCCC TGCACCTCCT GTCCTCTCCC AGTTCAATTCC

*Fig. 4A*

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2281 GGGTCACGCC TCCCACCCCG GCGCCCTCCC TGCACCTCCT GTCCTCTCCC AGTTCATTCC  
2341 TGGAACCAGC CAGGCCAGGC AACCAGTGGC CCCCAGAGG AGGCAGGATC CTCAGGCCCG  
2401 AGCCGCGGGA GGCTGGAAGG GCTGGCAGAT CGCTTCCCTC ATCCACCTCC ACCGGTCCAG  
2461 GTCTTTGCTG CTGTCCCAG ACCTCCTGTG ACACCAGGCC AGATCACAGG GCACCAGGCC  
2521 AGAGATAGTC TTCTTTTGT CCTTCTGGC CTCTGGCTAG TCAGTTTTTC ATAGCCTTAC  
2581 AGTATCTGGC TTTGTACTGA GAAATAAAC ACATTTTC

*Fig. 4B*

MALVTVSRSPPGSGASTPVGPWDQAVQRRSRLQRRQSFVLRGAVLGLQDGGDNDAAEASSEPTKAPSEELHGD  
QTDFGQGSQSPQKQEEQRQHLHLMVQLLRPQDDIRLAAQLEAPRPPRLRYLLVVSTREGEGLSQDETLLGVDFPDS  
SSPSCTLGLVPLWSDTQVYLDGDDGFSVTSGGQSRIFKPISTQTMWATLQVLHQACEAALGSLVPGGSALTWASH  
YQERLNSEQSCLNEWAMADLESLRPPSAEPGGSSEQMEQATRAELWKVLDVSDLESVTSKEIROALELRGLPL  
QQYRDFIDNQMLLLVAQRDRASRIFPHLYLGSEWNAANLELQRNRVTHILNMAREIDNFYPERFTYHNVRWDEES  
AQLLPHWKETHRFIEAARAQGHVVLVHCKMGVSRSAATVLAYAMKQYECSEALRHHVQLRPIARPNGFLRQLQI  
YQGILTART

*Fig. 5*

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1.....10.....20.....30.....40.....50.....60.....70.....80.....90.....100.....110.....120.....130  
 DSP-15 MALVTYSRPPSGASTPVGPMDQVQRSLQRQSFVAVLRGAV-LGLQDGGNDAAEASSETEKAPSEELHGGQTDGQSQSPQKQEFQRQHLHLMVQLLRPQDDIRLAQLEAPRPRLRYLL  
 AB036834 QSERRLSTDSTRSSNSTOSNDSIQHLQSMFYLLQREDTLKMAVKLESQPSNRTRVYLVIASRSCSGTSDRRRHRIMRHHSVKVGSAGTKSSTSPAVTQRQLSVEQTEATEASSCKDTADKENATA  
 DSP-13 MTLSTLARKRKAPLACTSLGGPDMTPYFSANAVISQAINQLISEFLTVKGAALFLPRNGSSTPRISSHRNKHA-----GDLQQLQAMFILLRPEDNIRLAVRLESTYQNRTRYWV  
 DSP-12  
 Consensus .....t.....s.....h.....l.....a.....  
 DSP-15 vvtstregelsgdetvlllgvdfpdsspsctlgvlplwsdtqvylgdggfsvtsggqsrifkpsiqtmwatlgvlgqacehalgsglvpggasalwshyqerlnseqsclnewtamadleslrpps  
 AB036834 AGDNKNTSGM--EESCLLGIDCNERT----TIGLWVPLADTTIHLDDGGFSVKVYEKTHIFKPVSVQAMMSALQTLHKYKSKARENWYASGSPSHDLSSYERRIESDQSCLEWNAWDALESRRPPS  
 dsp-13 WVSTNGRQDT--EESIVLGMDFSSNDSSTCTMGVLPLWSDTLIHLDDGGFSVSTONRVHIFKPVSVQAMMSALQSLHKACEVARAHNYPPGSLFLTWVSYESHINSDDQSSVNEWNAWQDVQSHRPS  
 dsp-12 MVLRLWSDTKIHLDDGGFSVSTAGRWIHFKPVSVQAMMSALQVLHKACEVARRHNYPPGVALIWATYYESCISSEQSCINEWNAWQDLESTRPDS  
 Consensus .....e...lg.d.....t.gslplusDT..hlDDGGFSVst.gr.hIFKPSIQAMMSALQVLHKaceAr.hny.pgg.al.WasyY#s.i.#QSc.NEWNAWQD1#S.RPds  
 DSP-15 rep--gsssegeqmeqairaelwkvldvsdlesvtскеirqalelriglpqayrfdi dnqmlLLVAQRORASRIFPHLYLGSEMNAAALEELQRNRVTHILMWAREIDNFYPERFTYHNVRLWDESAQ  
 ab036834 PDAIRNKPEKEETESVTKMKLKAIMSVLDDEVTSKYTRGRLEETLDMDLGEKSFDAEMLVILGQMDAPTKIFEHVYLGSEMNASNLEELQKNGVRHILNVTREIDNFFPGTFFYFNVRVYDDEKTN  
 dsp-13 PALFTDIPTERTERLTKTLREIMMQOLENTSKEIRTELEMQWCVNLREFKEFTDNEMIVILGQMSPTQIFEHVFLGSEMNASNLEELQKNGVRHILNVTREIDNFFPGTFFYFNVRVYDDEATD  
 dsp-12 PALFVDKPTGEGERTERLTKAKLRSTIMSQDLENVTSKEIRNELEKQWCVNLKELKEFTDNEMILLGQMDKPSLIFDHLVYLGSEMNASNLEELQSGVDVILNVTREIDNFFPGLFAYHNRVYDEETD  
 Consensus palff.dkpte.ErtEr1Ikaklr.!\$ms.DL#n!ITSKeIR.eLe.q\$.cml.#.kefIDn#m1li1gqmd.ps.IF.H1xLGSEMNASNLE#LQ..gv.yILNVtREIDNFxPg.F.Yhn!RvyD#e.t#  
 DSP-15 LLPHMKEHRETEAARAQSTHVLVHCKMGVSRSAATVIAVANKQYECSEALRHVQELRPIARPNGPFLRQLQIYQGIILTASRQSHWMEQKVGVSPEEHPAPEVSTFPPLPPEPEGGEKKEVGMEE  
 AB036834 LLKYWDDTFRYITRAKAESKVLVHCKMGVSRSAVIAVANKAYQMEFQALEHVKRSCIKPKNFNLQLETYSGLDAMKNKEKLQSKSEINLKSTKDARLLPGSEPTPLI-QALNQAKSSTGE  
 DSP-13 LLAYWNDTYKFTSKAKKHGSKCLVHCKMGVSRSAVIAVANKYQWNLDRAYDVKERRVTTPKPNPSFMRQLCEYQGIILLASFLGLTHGGDKDPWGEKSTEFESV--DLVSPGSPSCNPEKLLHISH  
 DSP-12 LLAHINEAYHFINKAKRNHCKCLVHCKMGVSRSAVIAVANKYQWNLDRAYDVKERRVTTPKPNPSFMRQLCEYQGIILLASFLGLTHGGDKDPWGEKSTEFESV--DLVSPGSPSCNPEKLLHISH  
 Consensus LLahn#cy.xI.kak..gskclVHCKMGVSRSAVIAVANKYQWNLDRAYDVKERRVTTPKPNPSFMRQLCEYQGIILLASFLGLTHGGDKDPWGEKSTEFESV--DLVSPGSPSCNPEKLLHISH

Fig. 6A

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521 530 540 550 560 570 580 590 600 610 620 630 640 650  
DSP-15 SOAPKEEPGRPRINLRGWMRSISLLEPSLELESTSETSDMPEVFSHSHSHEEPLQPFQLART-KGGQVDRGPQPAKSRQSVVTLOGSAVANRTOAFQEQGQGGQGGQGEPCISSTPRFKVVR  
AB036834 AGVTPDGEEDGSRWHRRSIAQKSQ--RRWRRSSSTSPKTQTAVVTQSQSMENLTPERSVAEEPKNMRFPGENSVTONQVLHIQKHTPLSVTRIHDL EAHRAQQLPQQPVWTSITKLITQTS  
DSP-13 PYLTPSIK  
DSP-12 PFLDDAQPGLGPPLPCCF-----RRLSDPLLSPDEAGSLVHLEDPEREALLEEAPPAEIVHRPARQDQG--SGLCEKDVKKLEFGSPKGRSGSLLOVEETEREEGLGAGRMGQLP-----T  
Consensus p.l.p...pg.gp.....rr.....s.....v.....e.l.....ae.....rqp.g...l.....l.....p.....q.ee.....g.g...w...lp.....  
651 660 670 680 690 700 710 720 730 740 746  
DSP-15 qasvhdsggeea  
AB036834 HLGKSVSGSSGNIDSRDSSCSDVFSSQVDSVFAKDEGEKRQRKTHSITESLGPSSGIVLDPTPQQKQSSNALRPRGTRQRELPSRHASMGs  
DSP-13  
DSP-12 QLDQNLNSENLMNSKR--SCPNI-----GWEVGRARPAGMHTPSLPSHS-----NWPTSASWGTGTGTRIHTQLIFFYCLLWAPSSHLQGPESFTG  
Consensus ql.....se..n..s.r..sc.....a...g.....hs.....p....v...t.....q.....l.....s....

*Fig. 6B*

## SEQUENCE LISTING

<110> Ceptyr Inc.  
Luche, Ralf M.  
Wei, Bo

<120> DSP-15 DUAL-SPECIFICITY PHOSPHATASE

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<140> PCT

<141> 2001-09-19

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<213> Homo sapiens

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ctccgtgggg ctgtcctggg actgcaggat ggaggggaca atgatgatgc agcagaggcc 180
agttctgagc caacagagaa ggccccgagt gaggaggagc tccacgggga ccagacagac 240
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<210> 2

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&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 2

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Thr Pro Val Gly Pro Trp Asp Gln Ala Val Gln Arg Arg Ser Arg Leu
      20      25      30
Gln Arg Arg Gln Ser Phe Ala Val Leu Arg Gly Ala Val Leu Gly Leu
      35      40      45
Gln Asp Gly Gly Asp Asn Asp Asp Ala Ala Glu Ala Ser Ser Glu Pro
      50      55      60
Thr Glu Lys Ala Pro Ser Glu Glu Glu Leu His Gly Asp Gln Thr Asp
      65      70      75      80
Phe Gly Gln Gly Ser Gln Ser Pro Gln Lys Gln Glu Glu Gln Arg Gln
      85      90      95
His Leu His Leu Met Val Gln Leu Leu Arg Pro Gln Asp Asp Ile Arg
      100      105      110
Leu Ala Ala Gln Leu Glu Ala Pro Arg Pro Pro Arg Leu Arg Tyr Leu
      115      120      125
Leu Val Val Ser Thr Arg Glu Gly Glu Gly Leu Ser Gln Asp Glu Thr
      130      135      140
Val Leu Leu Gly Val Asp Phe Pro Asp Ser Ser Ser Pro Ser Cys Thr
      145      150      155      160
Leu Gly Leu Val Leu Pro Leu Trp Ser Asp Thr Gln Val Tyr Leu Asp
      165      170      175
Gly Asp Gly Gly Phe Ser Val Thr Ser Gly Gly Gln Ser Arg Ile Phe
      180      185      190
Lys Pro Ile Ser Ile Gln Thr Met Trp Ala Thr Leu Gln Val Leu His
      195      200      205
Gln Ala Cys Glu Ala Ala Leu Gly Ser Gly Leu Val Pro Gly Gly Ser
      210      215      220
Ala Leu Thr Trp Ala Ser His Tyr Gln Glu Arg Leu Asn Ser Glu Gln
      225      230      235      240
Ser Cys Leu Asn Glu Trp Thr Ala Met Ala Asp Leu Glu Ser Leu Arg
      245      250      255
Pro Pro Ser Ala Glu Pro Gly Gly Ser Ser Glu Gln Glu Gln Met Glu
      260      265      270
Gln Ala Ile Arg Ala Glu Leu Trp Lys Val Leu Asp Val Ser Asp Leu
      275      280      285
Glu Ser Val Thr Ser Lys Glu Ile Arg Gln Ala Leu Glu Leu Arg Leu
      290      295      300
Gly Leu Pro Leu Gln Gln Tyr Arg Asp Phe Ile Asp Asn Gln Met Leu
      305      310      315      320
Leu Leu Val Ala Gln Arg Asp Arg Ala Ser Arg Ile Phe Pro His Leu
      325      330      335
Tyr Leu Gly Ser Glu Trp Asn Ala Ala Asn Leu Glu Glu Leu Gln Arg
      340      345      350
Asn Arg Val Thr His Ile Leu Asn Met Ala Arg Glu Ile Asp Asn Phe
      355      360      365
Tyr Pro Glu Arg Phe Thr Tyr His Asn Val Arg Leu Trp Asp Glu Glu
      370      375      380
Ser Ala Gln Leu Leu Pro His Trp Lys Glu Thr His Arg Phe Ile Glu
      385      390      395      400
Ala Ala Arg Ala Gln Gly Thr His Val Leu Val His Cys Lys Met Gly
      405      410      415
Val Ser Arg Ser Ala Ala Thr Val Leu Ala Tyr Ala Met Lys Gln Tyr
      420      425      430
Glu Cys Ser Leu Glu Gln Ala Leu Arg His Val Gln Glu Leu Arg Pro
      435      440      445
Ile Ala Arg Pro Asn Pro Gly Phe Leu Arg Gln Leu Gln Ile Tyr Gln
      450      455      460

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Gly Ile Leu Thr Ala Ser Arg Gln Ser His Val Trp Glu Gln Lys Val  
 465 470 475 480  
 Gly Gly Val Ser Pro Glu Glu His Pro Ala Pro Glu Val Ser Thr Pro  
 485 490 495  
 Phe Pro Pro Leu Pro Pro Glu Pro Glu Gly Gly Gly Glu Glu Lys Val  
 500 505 510  
 Val Gly Met Glu Glu Ser Gln Ala Pro Lys Glu Glu Pro Gly Pro  
 515 520 525  
 Arg Pro Arg Ile Asn Leu Arg Gly Val Met Arg Ser Ile Ser Leu Leu  
 530 535 540  
 Glu Pro Ser Leu Glu Leu Glu Ser Thr Ser Glu Thr Ser Asp Met Pro  
 545 550 555 560  
 Glu Val Phe Ser Ser His Glu Ser Ser His Glu Glu Pro Leu Gln Pro  
 565 570 575  
 Phe Pro Gln Leu Ala Arg Thr Lys Gly Gly Gln Gln Val Asp Arg Gly  
 580 585 590  
 Pro Gln Pro Ala Leu Lys Ser Arg Gln Ser Val Val Thr Leu Gln Gly  
 595 600 605  
 Ser Ala Val Val Ala Asn Arg Thr Gln Ala Phe Gln Glu Gln Glu Gln  
 610 615 620  
 Gly Gln Gly Gln Gly Gln Gly Glu Pro Cys Ile Ser Ser Thr Pro Arg  
 625 630 635 640  
 Phe Arg Lys Val Val Arg Gln Ala Ser Val His Asp Ser Gly Glu Glu  
 645 650 655  
 Gly Glu Ala

<210> 3  
 <211> 156  
 <212> PRT  
 <213> Homo sapiens

<400> 3  
 Asp Gly Ser Pro Leu Ser Asn Ser Gln Pro Ser Phe Pro Val Glu Ile  
 1 5 10 15  
 Leu Pro Phe Leu Tyr Leu Gly Cys Ala Lys Asp Ser Thr Asn Leu Asp  
 20 25 30  
 Val Leu Glu Glu Phe Gly Ile Lys Tyr Ile Leu Asn Val Thr Pro Asn  
 35 40 45  
 Leu Pro Asn Leu Phe Glu Asn Ala Gly Glu Phe Lys Tyr Lys Gln Ile  
 50 55 60  
 Pro Ile Ser Asp His Trp Ser Gln Asn Leu Ser Gln Phe Phe Pro Glu  
 65 70 75 80  
 Ala Ile Ser Phe Ile Asp Glu Ala Arg Gly Lys Asn Cys Gly Val Leu  
 85 90 95  
 Val His Cys Leu Ala Gly Ile Ser Arg Ser Val Thr Val Thr Val Ala  
 100 105 110  
 Tyr Leu Met Gln Lys Leu Asn Leu Ser Met Asn Asp Ala Tyr Asp Ile  
 115 120 125  
 Val Lys Met Lys Lys Ser Asn Ile Ser Pro Asn Phe Asn Phe Met Gly  
 130 135 140  
 Gln Leu Leu Asp Phe Glu Arg Thr Leu Gly Leu Ser  
 145 150 155

<210> 4  
 <211> 156  
 <212> PRT  
 <213> Homo sapiens

<400> 4

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Asp Gly Ser Pro Val Pro Ser Ser Gln Pro Ala Phe Pro Val Gln Ile
 1      5      10      15
Leu Pro Tyr Leu Tyr Leu Gly Cys Ala Lys Asp Ser Thr Asn Leu Asp
      20      25      30
Val Leu Gly Lys Tyr Gly Ile Lys Tyr Ile Leu Asn Val Thr Pro Asn
      35      40      45
Leu Pro Asn Ala Phe Glu His Gly Gly Glu Phe Thr Tyr Lys Gln Ile
      50      55      60
Pro Ile Ser Asp His Trp Ser Gln Asn Leu Ser Gln Phe Phe Pro Glu
65      70      75      80
Ala Ile Ser Phe Ile Asp Glu Ala Arg Ser Lys Lys Cys Gly Val Leu
      85      90      95
Val His Cys Leu Ala Gly Ile Ser Arg Ser Val Thr Val Thr Val Ala
      100      105      110
Tyr Leu Met Gln Lys Met Asn Leu Ser Leu Asn Asp Ala Tyr Asp Phe
      115      120      125
Val Lys Arg Lys Lys Ser Asn Ile Ser Pro Asn Phe Asn Phe Met Gly
130      135      140
Gln Leu Leu Asp Phe Glu Arg Thr Leu Gly Leu Ser
145      150      155

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<210> 5  
 <211> 156  
 <212> PRT  
 <213> Homo sapiens

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<400> 5
Ala Thr Pro Pro Pro Val Gly Leu Arg Ala Ser Phe Pro Val Gln Ile
 1      5      10      15
Leu Pro Asn Leu Tyr Leu Gly Ser Ala Arg Asp Ser Ala Asn Leu Glu
      20      25      30
Ser Leu Ala Lys Leu Gly Ile Arg Tyr Ile Leu Asn Val Thr Pro Asn
      35      40      45
Leu Pro Asn Phe Phe Glu Lys Asn Gly Asp Phe His Tyr Lys Gln Ile
      50      55      60
Pro Ile Ser Asp His Trp Ser Gln Asn Leu Ser Arg Phe Phe Pro Glu
65      70      75      80
Ala Ile Glu Phe Ile Asp Glu Ala Leu Ser Gln Asn Cys Gly Val Leu
      85      90      95
Val His Cys Leu Ala Gly Val Ser Arg Ser Val Thr Val Thr Val Ala
      100      105      110
Tyr Leu Met Gln Lys Leu His Leu Ser Leu Asn Asp Ala Tyr Asp Leu
      115      120      125
Val Lys Arg Lys Lys Ser Asn Ile Ser Pro Asn Phe Asn Phe Met Gly
130      135      140
Gln Leu Leu Asp Phe Glu Arg Ser Leu Arg Leu Glu
145      150      155

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<210> 6  
 <211> 155  
 <212> PRT  
 <213> Homo sapiens

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<400> 6
Leu Ser Gln Pro Cys Leu Pro Val Pro Ser Val Gly Leu Thr Arg Ile
 1      5      10      15
Leu Pro His Leu Tyr Leu Gly Ser Gln Lys Asp Val Leu Asn Lys Asp
      20      25      30
Leu Met Thr Gln Asn Gly Ile Ser Tyr Val Leu Asn Ala Ser Asn Ser
      35      40      45

```

Cys Pro Lys Pro Asp Phe Ile Cys Glu Ser Arg Phe Met Arg Val Pro  
 50 55 60  
 Ile Asn Asp Asn Tyr Cys Glu Lys Leu Leu Pro Trp Leu Asp Lys Ser  
 65 70 75 80  
 Ile Glu Phe Ile Asp Lys Ala Lys Leu Ser Ser Cys Gln Val Ile Val  
 85 90 95  
 His Cys Leu Ala Gly Ile Ser Arg Ser Ala Thr Ile Ala Ile Ala Tyr  
 100 105 110  
 Ile Met Lys Thr Met Gly Met Ser Ser Asp Asp Ala Tyr Arg Phe Val  
 115 120 125  
 Lys Asp Arg Arg Pro Ser Ile Ser Pro Asn Phe Asn Phe Leu Gly Gln  
 130 135 140  
 Leu Leu Glu Tyr Glu Arg Thr Leu Lys Leu Leu  
 145 150 155

<210> 7  
 <211> 154  
 <212> PRT  
 <213> Homo sapiens

<400> 7  
 Ser Asp Pro Arg Val Pro Ile Tyr Asp Gln Gly Gly Pro Val Glu Ile  
 1 5 10 15  
 Leu Pro Tyr Leu Tyr Leu Gly Ser Cys Asn His Ser Ser Asp Leu Gln  
 20 25 30  
 Gly Leu Gln Ala Cys Gly Ile Thr Ala Val Leu Asn Val Ser Ala Ser  
 35 40 45  
 Cys Pro Asn His Phe Glu Gly Leu Phe His Tyr Lys Ser Ile Pro Val  
 50 55 60  
 Glu Asp Asn Gln Met Val Glu Ile Ser Ala Trp Phe Gln Glu Ala Ile  
 65 70 75 80  
 Ser Phe Ile Asp Ser Val Lys Asn Ser Gly Gly Arg Val Leu Val His  
 85 90 95  
 Cys Gln Ala Gly Ile Ser Arg Ser Ala Thr Ile Cys Leu Ala Tyr Leu  
 100 105 110  
 Ile Gln Ser His Arg Val Arg Leu Asp Glu Ala Phe Asp Phe Val Lys  
 115 120 125  
 Gln Arg Arg Gly Val Ile Ser Pro Asn Phe Ser Phe Met Gly Gln Leu  
 130 135 140  
 Leu Gln Leu Glu Thr Gln Val Leu Cys His  
 145 150

<210> 8  
 <211> 154  
 <212> PRT  
 <213> Homo sapiens

<400> 8  
 Ser Ser Cys Ser Thr Pro Leu Tyr Asp Gln Gly Gly Pro Val Glu Ile  
 1 5 10 15  
 Leu Pro Phe Leu Tyr Leu Gly Ser Ala Tyr His Ala Ser Arg Lys Asp  
 20 25 30  
 Met Leu Asp Ala Leu Gly Ile Thr Ala Leu Ile Asn Val Ser Ala Asn  
 35 40 45  
 Cys Pro Asn His Phe Glu Gly His Tyr Gln Tyr Lys Ser Ile Pro Val  
 50 55 60  
 Glu Asp Asn His Lys Ala Asp Ile Ser Ser Trp Phe Asn Glu Ala Ile  
 65 70 75 80  
 Asp Phe Ile Asp Ser Ile Lys Asn Ala Gly Gly Arg Val Phe Val His  
 85 90 95

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Cys Gln Ala Gly Ile Ser Arg Ser Ala Thr Ile Cys Leu Ala Tyr Leu
      100      105
Met Arg Thr Asn Arg Val Lys Leu Asp Glu Ala Phe Glu Phe Val Lys
      115      120      125
Gln Arg Arg Ser Ile Ile Ser Pro Asn Phe Ser Phe Met Gly Gln Leu
      130      135      140
Leu Gln Phe Glu Ser Gln Val Leu Ala Pro
145      150

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<210> 9  
 <211> 154  
 <212> PRT  
 <213> Homo sapiens

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<400> 9
Ser Ser Cys Gly Thr Pro Leu His Asp Gln Gly Gly Pro Val Glu Ile
 1      5      10
Leu Pro Phe Leu Tyr Leu Gly Ser Ala Tyr His Ala Ala Arg Arg Asp
      20      25      30
Met Leu Asp Ala Leu Gly Ile Thr Ala Leu Leu Asn Val Ser Ser Asp
      35      40      45
Cys Pro Asn His Phe Glu Gly His Tyr Gln Tyr Lys Cys Ile Pro Val
 50      55      60
Glu Asp Asn His Lys Ala Asp Ile Ser Ser Trp Phe Met Glu Ala Ile
 65      70      75      80
Glu Tyr Ile Asp Ala Val Lys Asp Cys Arg Gly Arg Val Leu Val His
      85      90      95
Cys Gln Ala Gly Ile Ser Arg Ser Ala Thr Ile Cys Leu Ala Tyr Leu
      100      105      110
Met Met Lys Lys Arg Val Arg Leu Glu Glu Ala Phe Glu Phe Val Lys
      115      120      125
Gln Arg Arg Ser Ile Ile Ser Pro Asn Phe Ser Phe Met Gly Gln Leu
      130      135      140
Leu Gln Phe Glu Ser Gln Val Leu Ala Thr
145      150

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<210> 10  
 <211> 154  
 <212> PRT  
 <213> Homo sapiens

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<400> 10
Asn Val Ser Tyr Arg Pro Ala Tyr Asp Gln Gly Gly Pro Val Glu Ile
 1      5      10
Leu Pro Phe Leu Tyr Leu Gly Ser Ala Tyr His Ala Ser Lys Cys Glu
      20      25      30
Phe Leu Ala Asn Leu His Ile Thr Ala Leu Leu Asn Val Ser Arg Arg
      35      40      45
Thr Ser Glu Ala Cys Met Thr His Leu His Tyr Lys Trp Ile Pro Val
 50      55      60
Glu Asp Ser His Thr Ala Asp Ile Ser Ser His Phe Gln Glu Ala Ile
 65      70      75      80
Asp Phe Ile Asp Cys Val Arg Glu Lys Gly Gly Lys Val Leu Val His
      85      90      95
Cys Glu Ala Gly Ile Ser Arg Ser Pro Thr Ile Cys Met Ala Tyr Leu
      100      105      110
Met Lys Thr Lys Gln Phe Arg Leu Lys Glu Ala Phe Asp Tyr Ile Lys
      115      120      125
Gln Arg Arg Ser Met Val Ser Pro Asn Phe Gly Phe Met Gly Gln Leu
      130      135      140

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Leu Gln Tyr Glu Ser Glu Ile Leu Pro Ser  
145 150

<210> 11  
<211> 163  
<212> PRT  
<213> Homo sapiens

<400> 11  
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Thr Pro Arg Ile Tyr Val Gly Asn Ala Ser Val Ala Gln Asp Ile Pro  
20 25 30  
Lys Leu Gln Lys Leu Gly Ile Thr His Val Leu Asn Ala Ala Glu Gly  
35 40 45  
Arg Ser Phe Met His Val Asn Thr Asn Ala Asn Phe Tyr Lys Asp Ser  
50 55 60  
Gly Ile Thr Tyr Leu Gly Ile Lys Ala Asn Asp Thr Gln Glu Phe Asn  
65 70 75 80  
Leu Ser Ala Tyr Phe Glu Arg Ala Ala Asp Phe Ile Asp Gln Ala Leu  
85 90 95  
Ala Gln Lys Asn Gly Arg Val Leu Val His Cys Arg Glu Gly Tyr Ser  
100 105 110  
Arg Ser Pro Thr Leu Val Ile Ala Tyr Leu Met Met Arg Gln Lys Met  
115 120 125  
Asp Val Lys Ser Ala Leu Ser Ile Val Arg Gln Asn Arg Glu Ile Gly  
130 135 140  
Pro Asn Asp Gly Phe Leu Ala Gln Leu Cys Gln Leu Asn Asp Arg Leu  
145 150 155 160  
Ala Lys Glu

<210> 12  
<211> 140  
<212> PRT  
<213> Homo sapiens

<400> 12  
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1 5 10 15  
His Pro Ser Phe Ile Leu Asn Ser Ser Pro Ala His Ser Pro Met Ala  
20 25 30  
Arg Glu Ile Asp Asn Phe Tyr Pro Glu Arg Phe Thr Tyr His Asn Val  
35 40 45  
Arg Leu Trp Asp Glu Glu Ser Ala Gln Leu Leu Pro His Trp Lys Glu  
50 55 60  
Thr His Arg Phe Ile Glu Ala Ala Arg Ala Gln Gly Thr His Val Leu  
65 70 75 80  
Val His Cys Lys Met Gly Val Ser Arg Ser Ala Ala Thr Val Leu Ala  
85 90 95  
Tyr Ala Met Lys Gln Tyr Glu Cys Ser Leu Glu Gln Ala Leu Arg His  
100 105 110  
Val Gln Glu Leu Arg Pro Ile Ala Arg Pro Asn Pro Gly Phe Leu Arg  
115 120 125  
Gln Leu Gln Ile Tyr Gln Gly Ile Leu Thr Ala Arg  
130 135 140

<210> 13  
<211> 737

&lt;212&gt; PRT

&lt;213&gt; Drosophila melanogaster

&lt;400&gt; 13

Gln Ser Glu Arg Arg Leu Ser Thr Asp Ser Thr Arg Ser Ser Asn Ser  
 1 5 10 15  
 Thr Gln Ser Asn Asn Ser Asp Ile Gln Leu His Leu Gln Ser Met Phe  
 20 25 30  
 Tyr Leu Leu Gln Arg Glu Asp Thr Leu Lys Met Ala Val Lys Leu Glu  
 35 40 45  
 Ser Gln Arg Ser Asn Arg Thr Arg Tyr Leu Val Ile Ala Ser Arg Ser  
 50 55 60  
 Cys Cys Arg Ser Gly Thr Ser Asp Arg Arg His Arg Ile Met Arg  
 65 70 75 80  
 His His Ser Val Lys Val Gly Gly Ser Ala Gly Thr Lys Ser Ser Thr  
 85 90 95  
 Ser Pro Ala Val Pro Thr Gln Arg Gln Leu Ser Val Glu Gln Thr Ala  
 100 105 110  
 Thr Glu Ala Ser Ser Lys Cys Asp Lys Thr Ala Asp Lys Glu Asn Ala  
 115 120 125  
 Thr Ala Ala Gly Asp Asn Lys Asn Thr Ser Gly Met Glu Glu Ser Cys  
 130 135 140  
 Leu Leu Gly Ile Asp Cys Asn Glu Arg Thr Thr Ile Gly Leu Val Val  
 145 150 155 160  
 Pro Ile Leu Ala Asp Thr Thr Ile His Leu Asp Gly Asp Gly Gly Phe  
 165 170 175  
 Ser Val Lys Val Tyr Glu Lys Thr His Ile Phe Lys Pro Val Ser Val  
 180 185 190  
 Gln Ala Met Trp Ser Ala Leu Gln Thr Leu His Lys Val Ser Lys Lys  
 195 200 205  
 Ala Arg Glu Asn Asn Phe Tyr Ala Ser Gly Pro Ser His Asp Trp Leu  
 210 215 220  
 Ser Ser Tyr Glu Arg Arg Ile Glu Ser Asp Gln Ser Cys Leu Asn Glu  
 225 230 235 240  
 Trp Asn Ala Met Asp Ala Leu Glu Ser Arg Arg Pro Pro Ser Pro Asp  
 245 250 255  
 Ala Ile Arg Asn Lys Pro Pro Glu Lys Glu Glu Thr Glu Ser Val Ile  
 260 265 270  
 Lys Met Lys Leu Lys Ala Ile Met Met Ser Val Asp Leu Asp Glu Val  
 275 280 285  
 Thr Ser Lys Tyr Ile Arg Gly Arg Leu Glu Glu Ile Leu Asp Met Asp  
 290 295 300  
 Leu Gly Glu Tyr Lys Ser Phe Ile Asp Ala Glu Met Leu Val Ile Leu  
 305 310 315 320  
 Gly Gln Met Asp Ala Pro Thr Lys Ile Phe Glu His Val Tyr Leu Gly  
 325 330 335  
 Ser Glu Trp Asn Ala Ser Asn Leu Glu Glu Leu Gln Lys Asn Gly Val  
 340 345 350  
 Arg His Ile Leu Asn Val Thr Arg Glu Ile Asp Asn Phe Phe Pro Gly  
 355 360 365  
 Thr Phe Glu Tyr Phe Asn Val Arg Val Tyr Asp Asp Glu Lys Thr Asn  
 370 375 380  
 Leu Leu Lys Tyr Trp Asp Asp Thr Phe Arg Tyr Ile Thr Arg Ala Lys  
 385 390 395 400  
 Ala Glu Gly Ser Lys Val Leu Val His Cys Lys Met Gly Val Ser Arg  
 405 410 415  
 Ser Ala Ser Val Val Ile Ala Tyr Ala Met Lys Ala Tyr Gln Trp Glu  
 420 425 430  
 Phe Gln Gln Ala Leu Glu His Val Lys Lys Arg Arg Ser Cys Ile Lys  
 435 440 445  
 Pro Asn Lys Asn Phe Leu Asn Gln Leu Glu Thr Tyr Ser Gly Met Leu  
 450 455 460

Asp Ala Met Lys Asn Lys Glu Lys Leu Gln Arg Ser Lys Ser Glu Thr  
 465 470 475 480  
 Asn Leu Lys Ser Thr Lys Asp Ala Arg Leu Leu Pro Gly Ser Glu Pro  
 485 490 495  
 Thr Pro Leu Ile Gln Ala Leu Asn Gln Ala Lys Ser Lys Ser Thr Gly  
 500 505 510  
 Glu Ala Gly Val Thr Pro Asp Gly Glu Glu Glu Asp Gly Ser Arg Met  
 515 520 525  
 His Arg Arg Ser Ile Ala Gln Lys Ser Gln Arg Arg Met Val Arg Arg  
 530 535 540  
 Ser Ser Ser Thr Ser Pro Lys Thr Gln Thr Ala Val Val Thr Lys Gln  
 545 550 555 560  
 Gln Ser Gln Ser Met Glu Asn Leu Thr Pro Glu Arg Ser Val Ala Glu  
 565 570 575  
 Glu Pro Lys Asn Met Arg Phe Pro Gly Ser Asn Gly Glu Asn Tyr Ser  
 580 585 590  
 Val Thr Gln Asn Gln Val Leu His Ile Gln Lys His Thr Pro Leu Ser  
 595 600 605  
 Val Arg Thr Arg Ile His Asp Leu Glu Ala His Arg Ala Asp Gln Leu  
 610 615 620  
 Pro Gln Gln Pro Val Trp Thr Ser Leu Thr Lys Leu Ile Thr Gln Thr  
 625 630 635 640  
 Ser His Leu Gly Lys Ser Val Ser Gly Ser Ser Ser Gly Asn Ile Asp  
 645 650 655  
 Ser Arg Arg Asp Ser Ser Cys Ser Asp Val Phe Ser Ser Gln Val Asp  
 660 665 670  
 Ser Val Phe Ala Lys Asp Glu Gly Glu Lys Arg Gln Arg Arg Lys Thr  
 675 680 685  
 His Ser Trp Thr Glu Ser Leu Gly Pro Ser Gly Gly Ile Val Leu Asp  
 690 695 700  
 Pro Thr Pro Gln Gln Gln Lys Gln Gln Ser Asn Ala Ile Leu Arg Pro  
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 Arg Gly Thr Arg Gln Arg Glu Leu Pro Ser Arg His Ala Ser Trp Gly  
 725 730 735  
 Ser

<210> 14  
 <211> 509  
 <212> PRT  
 <213> Homo sapiens

<400> 14  
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 Ala Val Ile Ser Gln Asn Ala Ile Asn Gln Leu Ile Ser Glu Ser Phe  
 35 40 45  
 Leu Thr Val Lys Gly Ala Ala Leu Phe Leu Pro Arg Gly Asn Gly Ser  
 50 55 60  
 Ser Thr Pro Arg Ile Ser His Arg Arg Asn Lys His Ala Gly Asp Leu  
 65 70 75 80  
 Gln Gln His Leu Gln Ala Met Phe Ile Leu Leu Arg Pro Glu Asp Asn  
 85 90 95  
 Ile Arg Leu Ala Val Arg Leu Glu Ser Thr Tyr Gln Asn Arg Thr Arg  
 100 105 110  
 Tyr Met Val Val Val Ser Thr Asn Gly Arg Gln Asp Thr Glu Glu Ser  
 115 120 125  
 Ile Val Leu Gly Met Asp Phe Ser Ser Asn Asp Ser Ser Thr Cys Thr  
 130 135 140

Met Gly Leu Val Leu Pro Leu Trp Ser Asp Thr Leu Ile His Leu Asp  
 145 150 155 160  
 Gly Asp Gly Gly Phe Ser Val Ser Thr Asp Asn Arg Val His Ile Phe  
 165 170 175  
 Lys Pro Val Ser Val Gln Ala Met Trp Ser Ala Leu Gln Ser Leu His  
 180 185 190  
 Lys Ala Cys Glu Val Ala Arg Ala His Asn Tyr Tyr Pro Gly Ser Leu  
 195 200 205  
 Phe Leu Thr Trp Val Ser Tyr Tyr Glu Ser His Ile Asn Ser Asp Gln  
 210 215 220  
 Ser Ser Val Asn Glu Trp Asn Ala Met Gln Asp Val Gln Ser His Arg  
 225 230 235 240  
 Pro Asp Ser Pro Ala Leu Phe Thr Asp Ile Pro Thr Glu Arg Glu Arg  
 245 250 255  
 Thr Glu Arg Leu Ile Lys Thr Lys Leu Arg Glu Ile Met Met Gln Lys  
 260 265 270  
 Asp Leu Glu Asn Ile Thr Ser Lys Glu Ile Arg Thr Glu Leu Glu Met  
 275 280 285  
 Gln Met Val Cys Asn Leu Arg Glu Phe Lys Glu Phe Ile Asp Asn Glu  
 290 295 300  
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 305 310 315 320  
 His Val Phe Leu Gly Ser Glu Trp Asn Ala Ser Asn Leu Glu Asp Leu  
 325 330 335  
 Gln Asn Arg Gly Val Arg Tyr Ile Leu Asn Val Thr Arg Glu Ile Asp  
 340 345 350  
 Asn Phe Phe Pro Gly Val Phe Glu Tyr His Asn Ile Arg Val Tyr Asp  
 355 360 365  
 Glu Glu Ala Thr Asp Leu Leu Ala Tyr Trp Asn Asp Thr Tyr Lys Phe  
 370 375 380  
 Ile Ser Lys Ala Lys Lys His Gly Ser Lys Cys Leu Val His Cys Lys  
 385 390 395 400  
 Met Gly Val Ser Arg Ser Ala Ser Thr Val Ile Ala Tyr Ala Met Lys  
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 Glu Tyr Gly Trp Asn Leu Asp Arg Ala Tyr Asp Tyr Val Lys Glu Arg  
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 Arg Thr Val Thr Lys Pro Asn Pro Ser Phe Met Arg Gln Leu Glu Glu  
 435 440 445  
 Tyr Gln Gly Ile Leu Leu Ala Ser Phe Leu Gly Leu Ile His Gly Gly  
 450 455 460  
 Arg Asp Lys Pro Trp Gly Glu Lys Ser Thr Glu Phe Glu Ser Val Asp  
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<210> 15  
 <211> 55.  
 <212> PRT  
 <213> Homo sapiens

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 35 40 45  
 Cys Glu Val Ala Arg Arg His Asn Tyr Phe Pro Gly Gly Val Ala Leu  
 50 55 60



Ile	Trp	Ala	Thr	Tyr	Tyr	Glu	Ser	Cys	Ile	Ser	Ser	Glu	Gln	Ser	Cys
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Ser	Pro	Ala	Leu	Phe	Val	Asp	Lys	Pro	Thr	Glu	Gly	Glu	Arg	Thr	Glu
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Arg	Leu	Ile	Lys	Ala	Lys	Leu	Arg	Ser	Ile	Met	Met	Ser	Gln	Asp	Leu
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Asn	Cys	Asn	Leu	Lys	Glu	Leu	Lys	Glu	Phe	Ile	Asp	Asn	Glu	Met	Leu
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Leu	Ile	Leu	Gly	Gln	Met	Asp	Lys	Pro	Ser	Leu	Ile	Phe	Asp	His	Leu
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Tyr	Leu	Gly	Ser	Glu	Trp	Asn	Ala	Ser	Asn	Leu	Glu	Glu	Leu	Gln	Gly
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Ser	Gly	Val	Asp	Tyr	Ile	Leu	Asn	Val	Thr	Arg	Glu	Ile	Asp	Asn	Phe
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Phe	Pro	Gly	Leu	Phe	Ala	Tyr	His	Asn	Ile	Arg	Val	Tyr	Asp	Glu	Glu
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Thr	Thr	Asp	Leu	Leu	Ala	His	Trp	Asn	Glu	Ala	Tyr	His	Phe	Ile	Asn
225					230					235					240
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Val	Ser	Arg	Ser	Ala	Ser	Thr	Val	Ile	Ala	Tyr	Ala	Met	Lys	Glu	Phe
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Gly	Trp	Pro	Leu	Glu	Lys	Ala	Tyr	Asn	Tyr	Val	Lys	Gln	Lys	Arg	Ser
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Gln	Thr	Asp	Ser	Ser	Leu	Gln	Gln	Pro	Val	Asp	Asp	Pro	Ala	Gly	Pro
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Cys	Cys	Phe	Arg	Arg	Leu	Ser	Asp	Pro	Leu	Leu	Pro	Ser	Pro	Glu	Asp
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Glu	Thr	Gly	Ser	Leu	Val	His	Leu	Glu	Asp	Pro	Glu	Arg	Glu	Ala	Leu
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Leu	Glu	Glu	Ala	Ala	Pro	Pro	Ala	Glu	Val	His	Arg	Pro	Ala	Arg	Gln
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Glu	Phe	Gly	Ser	Pro	Lys	Gly	Arg	Ser	Gly	Ser	Leu	Leu	Gln	Val	Glu
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Glu	Thr	Glu	Arg	Glu	Glu	Gly	Leu	Gly	Ala	Gly	Arg	Trp	Gly	Gln	Leu
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Asn	Ser	Lys	Arg	Ser	Cys	Pro	Asn	Gly	Met	Glu	Val	Gly	Arg	Ala	Arg
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Pro	Ala	Gly	Trp	His	Thr	Pro	Ser	Leu	Pro	Ser	His	Ser	Asn	Trp	Pro
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Thr	Ser	Ala	Ser	Val	Val	Gly	Thr	Thr	Gly	Thr	Arg	His	His	Thr	Gln
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Leu	Ile	Phe	Phe	Tyr	Cys	Leu	Leu	Trp	Ala	Pro	Ser	Ser	His	Leu	Gln
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545					550										

<210> 16  
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 <212> PRT  
 <213> Homo sapiens

<400> 16  
 Val His Cys Lys Met Gly Val Ser Arg Ser  
 1 5 10

<210> 17  
 <211> 24  
 <212> PRT  
 <213> Artificial Sequence

<220>  
 <223> Conserved homology region from eight DSPs having  
 MAP-kinase phosphatase activity

<400> 17  
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<210> 18  
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 <213> Homo sapiens

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<210> 19  
 <211> 30  
 <212> DNA  
 <213> Artificial Sequence

<220>  
 <223> Primer

<400> 19  
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<210> 20  
 <211> 1416  
 <212> DNA  
 <213> Mus musculus

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 ctccgtgggg ctgtcctggg actgcaggat ggaggggaca atgatgatgc agcagaggcc 180  
 agttctgagc caacagagaa ggccccgagt gaggaggagc tccacgggga ccagacagac 240  
 ttcgggcaag gatcccagag tcccagaag caggaggagc agaggcagca cctgcacctc 300

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caggatgaga cggctcctct gggcgtggat ttccctgaca gcagctcccc cagctgcacc 480
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ttcagcgtga cgtctggtgg gcaaagccgg atcttcaagc ccattctccat ccagaccatg 600
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 <213> Mus musculus

<400> 21

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Gln	Arg	Arg	Gln	Ser	Phe	Ala	Val	Leu	Arg	Gly	Ala	Val	Leu	Gly	Leu
			35				40					45			
Gln	Asp	Gly	Gly	Asp	Asn	Asp	Asp	Ala	Ala	Glu	Ala	Ser	Ser	Glu	Pro
	50				55					60					
Thr	Glu	Lys	Ala	Pro	Ser	Glu	Glu	Glu	Leu	His	Gly	Asp	Gln	Thr	Asp
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Phe	Gly	Gln	Gly	Ser	Gln	Ser	Pro	Gln	Lys	Gln	Glu	Glu	Gln	Arg	Gln
				85				90						95	
His	Leu	His	Leu	Met	Val	Gln	Leu	Leu	Arg	Pro	Gln	Asp	Asp	Ile	Arg
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Gly	Asp	Gly	Gly	Phe	Ser	Val	Thr	Ser	Gly	Gly	Gln	Ser	Arg	Ile	Phe
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Ala	Leu	Thr	Trp	Ala	Ser	His	Tyr	Gln	Glu	Arg	Leu	Asn	Ser	Glu	Gln
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Ser	Cys	Leu	Asn	Glu	Trp	Thr	Ala	Met	Ala	Asp	Leu	Glu	Ser	Leu	Arg
				245					250					255	
Pro	Pro	Ser	Ala	Glu	Pro	Gly	Gly	Ser	Ser	Glu	Gln	Glu	Gln	Met	Glu
			260					265					270		
Gln	Ala	Ile	Arg	Ala	Glu	Leu	Trp	Lys	Val	Leu	Asp	Val	Ser	Asp	Leu
			275				280					285			

Glu Ser Val Thr Ser Lys Glu Ile Arg Gln Ala Leu Glu Leu Arg Leu  
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 Gly Leu Pro Leu Gln Gln Tyr Arg Asp Phe Ile Asp Asn Gln Met Leu  
 305 310 315  
 Leu Leu Val Ala Gln Arg Asp Arg Ala Ser Arg Ile Phe Pro His Leu  
 325 330 335  
 Tyr Leu Gly Ser Glu Trp Asn Ala Ala Asn Leu Glu Glu Leu Gln Arg  
 340 345 350  
 Asn Arg Val Thr His Ile Leu Asn Met Ala Arg Glu Ile Asp Asn Phe  
 355 360 365  
 Tyr Pro Glu Arg Phe Thr Tyr His Asn Val Arg Leu Trp Asp Glu Glu  
 370 375 380  
 Ser Ala Gln Leu Leu Pro His Trp Lys Glu Thr His Arg Phe Ile Glu  
 385 390 395 400  
 Ala Ala Arg Ala Gln Gly Thr His Val Leu Val His Cys Lys Met Gly  
 405 410 415  
 Val Ser Arg Ser Ala Ala Thr Val Leu Ala Tyr Ala Met Lys Gln Tyr  
 420 425 430  
 Glu Cys Ser Leu Glu Gln Ala Leu Arg His Val Gln Glu Leu Arg Pro  
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 Gly Ile Leu Thr Ala Arg Thr  
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<210> 22  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence

<220>  
 <223> Primer

<400> 22  
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24

<210> 23  
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 <212> DNA  
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 <223> Primer

<400> 23  
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27

<210> 24  
 <211> 28  
 <212> DNA  
 <213> Artificial Sequence

<220>  
 <223> Primer

<400> 24  
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28

<210> 25  
 <211> 27  
 <212> DNA

15

BEST AVAILABLE COPY

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Primer

&lt;400&gt; 25

cgcagagact ccaggtcggc catagcc

27

&lt;210&gt; 26

&lt;211&gt; 23

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Primer

&lt;400&gt; 26

gggggttgagg gaaggggccg tgc

23

&lt;210&gt; 27

&lt;211&gt; 6

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 27

Asp Ala Asp Glu Tyr Leu

1

5

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